

## Overview

### Useful For

Prenatal testing for hemophilia A when a variant has not been identified in the family

### Reflex Tests

Test Id	Reporting Name	Available Separately	Always Performed
CULAF	Amniotic Fluid Culture/Genetic Test	Yes	No
CULFB	Fibroblast Culture for Genetic Test	Yes	No
MATCC	Maternal Cell Contamination, B	Yes	No
_STR1	Comp Analysis using STR (Bill only)	No, (Bill only)	No
_STR2	Add'l comp analysis w/STR (Bill Only)	No, (Bill only)	No

### Genetics Test Information

This test detects the common inversion variants within the *F8* gene. Approximately 50% of affected male patients with severe hemophilia A have been shown to have an inversion.

It is recommended that the *F8* inversion variant be confirmed in the affected male patient or obligate female carrier prior to testing at-risk individuals.

### Testing Algorithm

If amniotic fluid is received, amniotic fluid culture for genetic testing will be added at an additional charge.

If chorionic villus specimen is received, fibroblast culture for genetic testing will be added at an additional charge.

For any prenatal specimen that is received, maternal cell contamination studies will be added. A maternal whole blood specimen **is required** to perform this test.

The following algorithms are available:

-[Hemophilia Carrier Testing Algorithm](#)

-[Hemophilia Testing Algorithm](#)

### Special Instructions

- [Informed Consent for Genetic Testing](#)
- [Hemophilia Carrier Testing Algorithm](#)
- [Hemophilia Testing Algorithm](#)
- [Hemophilia A Patient Information](#)
- [Informed Consent for Genetic Testing \(Spanish\)](#)

**Method Name**

Polymerase Chain Reaction (PCR) or Inverse Shifting-Polymerase Chain Reaction (IS-PCR)

**NY State Available**

Yes

**Specimen****Specimen Type**

Varies

**Additional Testing Requirements**

Due to its complexity, consultation with the laboratory is required for all prenatal testing; call 800-533-1710 to speak to a genetic counselor.

**All prenatal specimens must be accompanied by a maternal blood specimen.** Order MATCC / Maternal Cell Contamination, Molecular Analysis, Varies on the maternal specimen.

**Shipping Instructions**

Advise Express Mail or equivalent if not on courier service.

**Necessary Information**

**[Hemophilia A Patient Information \(T712\)](#) is required.** Testing may proceed without the patient information, however, the information aids in providing a more thorough interpretation. Ordering providers are strongly encouraged to fill out the form and send with the specimen.

**Specimen Required**

Results will be reported and telephoned or faxed if requested.

**Submit only 1 of the following specimens:**

**Specimen Type:** Amniotic fluid

**Container/Tube:** Amniotic fluid container

**Specimen Volume:** 5-10 mL

**Collection Instructions:**

1. Optimal timing for specimen collection is during 14 to 18 weeks of gestation, but specimens collected at other weeks of gestation are also accepted.
2. Discard the first 2 mL of amniotic fluid. If the culture will be performed in conjunction with chromosome analysis and alpha-fetoprotein, a total of approximately 25 mL to 30 mL will be needed for the combined studies.

**Specimen Stability Information:** Ambient (preferred) <24 hours/Refrigerated

**Additional Information:**

1. Place the tubes in a Styrofoam container.
2. Fill remaining space with packing material.

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3. Unavoidably, about 1% to 2% of mailed-in specimens are not viable.
4. Bloody specimens are undesirable.
5. If the specimen does not grow in culture, you will be notified within 7 days of receipt.
6. A separate culture charge will be assessed under CULAF / Culture for Genetic Testing, Amniotic Fluid.
- 7. All prenatal specimens must be accompanied by a maternal blood specimen;** order MATCC / Maternal Cell Contamination, Molecular Analysis, Varies on the maternal specimen.

**Specimen Type:** Chorionic villi

**Supplies:** CVS Media (RPMI) and Small Dish (T095)

**Container/Tube:** 15-mL tube containing 15 mL of transport media

**Specimen Volume:** 20-30 mg

**Collection Instructions:**

1. Collect specimen by the transabdominal or transcervical method.
2. Transfer the chorionic villi specimen to a Petri dish containing transport medium.
3. Using a stereomicroscope and sterile forceps, assess the quality and quantity of the villi and remove any blood clots and maternal decidua.

**Specimen Stability Information:** Refrigerated (preferred) <24 hours/Ambient

**Additional Information:**

1. A separate culture charge will be assessed under CULFB / Fibroblast Culture for Biochemical or Molecular Testing. An additional 3 to 4 weeks is required to culture fibroblasts before genetic testing can occur.
- 2. All prenatal specimens must be accompanied by a maternal blood specimen;** order MATCC / Maternal Cell Contamination, Molecular Analysis, Varies on the maternal specimen.

**Specimen Type:** Confluent cultured cells

**Container/Tube:** T-25 flask

**Specimen Volume:** 2 Flasks approximately 90% confluent

**Collection Instructions:** Submit confluent cultured cells from another laboratory

**Specimen Stability Information:** Ambient (preferred) <24 hours/Refrigerated

**Additional Information:** [All prenatal specimens must be accompanied by a maternal blood specimen;](#) order MATCC / Maternal Cell Contamination, Molecular Analysis, Varies on the maternal specimen.

**Forms**

- [1. Hemophilia A Patient Information \(T712\) is required.](#)
- 2. New York Clients-Informed consent is required.** Document on the request form or electronic order that a copy is on file. The following documents are available:  
-[Informed Consent for Genetic Testing](#) (T576)  
-[Informed Consent for Genetic Testing-Spanish](#) (T826)
3. If not ordering electronically, complete, print, and send a [Coagulation Test Request](#) (T753) with the specimen.

**Specimen Minimum Volume**

Amniotic fluid: See Specimen Required

Chorionic villi: 5 mg

**Reject Due To**

All specimens will be evaluated at Mayo Clinic Laboratories for test suitability.

**Specimen Stability Information**

Specimen Type	Temperature	Time	Special Container
Varies	Varies		

**Clinical & Interpretive****Clinical Information**

Hemophilia A (HA) is due to a deficiency of clotting factor VIII (FVIII). HA is an X-linked recessive bleeding disorder that affects approximately 1 in 5000 male individuals. Male patients are typically affected with bleeding symptoms, whereas female carriers generally do not have bleeding symptoms but are at risk of having affected sons. Rarely, approximately 10% of female carriers have FVIII activity levels below 35% and are at risk for bleeding.

Bleeding, the most common clinical symptom in individuals with HA, correlates with FVIII activity levels. FVIII activity levels of below 1% are associated with severe disease, 1% to 5% activity with moderate disease, and 5% to 40% with mild disease. In male patients with severe deficiency, spontaneous bleeding may occur. In individuals with mild HA, bleeding may occur only after surgery or trauma.

FVIII is encoded by the factor VIII (*F8*) gene. Approximately 98% of patients with a diagnosis of HA are found to have a variant in *F8* (ie, intron 1 and 22 inversions, point mutations, insertions, and deletions). The intron 1 and 22 inversion variants account for approximately 50% of variants associated with severe HA. These inversions are typically not identified in patients with mild or moderate HA.

It is recommended that the *F8* variant be confirmed in the affected male patient or obligate female carrier prior to testing at-risk individuals. Affected male patients are identified by FVIII activity (F8A / Coagulation Factor VIII Activity Assay, Plasma) and clinical evaluation, while obligate female carriers are identified by family history assessment. If the intron inversion assays do not detect an inversion in these individuals, additional analysis (ie, *F8* sequencing) may be able to identify the familial variant. Of note, not all women with an affected son are germline carriers of a *F8* variant, as *de novo* variants in *F8* do occur. Approximately 20% of mothers of isolated cases do not have an identifiable germline *F8* variant. Importantly, there is a small risk for recurrence even when the familial *F8* variant is not identified in the mother of the affected patient due to the possibility of germline mosaicism.

**Reference Values**

An interpretive report will be provided.

**Interpretation**

The interpretive report will include assay information, background information, and conclusions based on the test results.

**Cautions**

Obtaining a medical genetics or hematology (coagulation) consultation prior to ordering is advisable. Molecular genetic or hemophilia center consultation is available for all possible hemophilia A cases and is particularly indicated in complex cases or in situations in which the diagnosis is atypical or uncertain.

This assay detects only *F8* intron 1 and 22 inversion variants. Thus, a negative result does not exclude the presence of other variants in *F8*.

The intron 1 and 22 inversion variants targeted by this assay are found in approximately 50% of individuals with severe hemophilia A; the assay may be uninformative for a number of families.

Test results should be interpreted in the context of clinical findings, family history, and other laboratory data. Errors in the interpretation of results may occur if the information provided is inaccurate or incomplete.

### **Clinical Reference**

1. Antonarakis SE, Rossiter JP, Young M, et al: Factor VIII gene inversions in severe hemophilia A: results of an international consortium study. *Blood*. 1995 Sep 15;86(6):2206-2212
2. Rossiter JP, Young M, Kimberland ML, et al: Factor VIII gene inversions causing severe hemophilia A originate almost exclusively in male germ cells. *Hum Mol Genet*. 1994 Jul;3(7):1035-1039
3. Castaldo G, D'Argenio V, Nardiello P, et al: Haemophilia A: molecular insights. *Clin Chem Lab Med*. 2007;45(4):450-461
4. Oldenburg J, Rost S, El-Maari O, et al: De novo factor VIII gene intron 22 inversion in a female carrier presents as a somatic mosaicism. *Blood*. 2000 Oct 15;96(8):2905-2906
5. Johnsen JM, Fletcher SN, Huston H, et al: Novel approach to genetic analysis and results in 3000 hemophilia patients enrolled in the My Life, Our Future initiative. *Blood Adv*. 2017 May 18;1(13):824-834  
doi:10.1182/bloodadvances.2016002923
6. Pruthi RK: Hemophilia: a practical approach to genetic testing. *Mayo Clin Proc*. 2005 Nov;80(11):1485-1499

### **Performance**

### **Method Description**

Genomic DNA from whole blood or cord blood is digested with restriction enzyme, ligated with T4 DNA ligase, and amplified by polymerase chain reaction (PCR) with primers specific for the *F8* intron 22 inversion variants.(Rosetti LC, Radic CP, Larripa IB, De Brasi CD: Developing a new generation of tests for genotyping hemophilia-causative rearrangements involving int22h and int1h hotspots in the factor VIII gene. *J Thromb Haemost*. 2008 May;6(5):830-836)

Genomic DNA from whole blood or cord blood is amplified by PCR with primers specific for the *F8* intron 1 inversion mutation.(Bagnall RD, Waseem N, Green PM, Giannelli F: Recurrent inversion breaking intron 1 of the factor VIII gene is a frequent cause of severe hemophilia A. *Blood*. 2002 Jan 1;99[1]:168-174)

### **PDF Report**

No

### **Day(s) Performed**

Monday through Friday

### **Report Available**

28 to 35 days

**Specimen Retention Time**

Extracted DNA: Indefinitely, from New York State: 90 days

**Performing Laboratory Location**

Mayo Clinic Laboratories - Rochester Main Campus

**Fees & Codes****Fees**

- Authorized users can sign in to [Test Prices](#) for detailed fee information.
- Clients without access to Test Prices can contact [Customer Service](#) 24 hours a day, seven days a week.
- Prospective clients should contact their account representative. For assistance, contact [Customer Service](#).

**Test Classification**

This test was developed and its performance characteristics determined by Mayo Clinic in a manner consistent with CLIA requirements. It has not been cleared or approved by the US Food and Drug Administration.

**CPT Code Information**

81403

**LOINC® Information**

Test ID	Test Order Name	Order LOINC® Value
F8INP	HA F8 Int 1/22 Inversion, AF or CVS	82343-5

Result ID	Test Result Name	Result LOINC® Value
35161	HA F8 Int 1/22 Reason for Referral	42349-1
35162	HA F8 Int 1/22 Inversion, AF or CVS	82343-5
35163	F8INP Interpretation	69047-9
35164	HA F8 Intron 1/22 Reviewed By	18771-6