

Overview

Useful For

Evaluating for the following in an algorithmic process for the HAEV1 / Hemolytic Anemia Evaluation, Blood; HBEL1 / Hemoglobin Electrophoresis Evaluation, Blood; MEV1 / Methemoglobinemia Evaluation, Blood; REVE2 / Erythrocytosis Evaluation, Blood; THEV1 / Thalassemia and Hemoglobinopathy Evaluation, Blood and Serum:

- Diagnosing of beta thalassemia intermedia or major
- Identifying a specific beta thalassemia sequence variant (ie, unusually severe beta thalassemia trait)
- Evaluating an abnormal hemoglobin electrophoresis identifying a rare beta-globin variant
- Evaluating chronic hemolytic anemia of unknown etiology
- Evaluating hereditary erythrocytosis with left-shifted p50 oxygen dissociation results
- Preconception screening when there is a concern for a beta-hemoglobin disorder based on family history

Method Name

Only orderable as a reflex. For more information see:

- HAEV1 / Hemolytic Anemia Evaluation, Blood
- HBEL1 / Hemoglobin Electrophoresis Evaluation, Blood
- MEV1 / Methemoglobinemia Evaluation, Blood
- REVE2 / Erythrocytosis Evaluation, Blood
- THEV1 / Thalassemia and Hemoglobinopathy Evaluation, Blood and Serum

Polymerase Chain Reaction (PCR)/Sanger Sequencing

NY State Available

Yes

Specimen

Specimen Type

Whole Blood EDTA

Specimen Required

Only orderable as a reflex. For more information see:

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- HBEL1 / Hemoglobin Electrophoresis Evaluation, Blood
- MEV1 / Methemoglobinemia Evaluation, Blood
- REVE2 / Erythrocytosis Evaluation, Blood
- THEV1 / Thalassemia and Hemoglobinopathy Evaluation, Blood and Serum

Specimen Type: Whole blood

Container/Tube:

Preferred: Lavender top (EDTA)

Acceptable: Yellow top (ACD)

Specimen Volume: 4 mL

Specimen Minimum Volume

1 mL

Reject Due To

Gross hemolysis	OK
Moderately to severely clotted	Reject

Specimen Stability Information

Specimen Type	Temperature	Time	Special Container
Whole Blood EDTA	Refrigerated	14 days	

Clinical & Interpretive

Clinical Information

Beta-globin gene sequencing is useful in the evaluation of beta-globin chain variants and beta thalassemia. It detects almost all beta-globin variants and the most common beta-thalassemia sequence variants, although prevalence is ethnicity dependent. Because these conditions are often complex, this test should always be interpreted in the context of protein studies, such as hemoglobin electrophoresis and red blood cell indices.

The majority of beta-globin chain variants are clinically and hematologically benign; however, some have important clinical consequences, such as erythrocytosis, cyanosis/hypoxia, chronic hemolysis, or unexplained microcytosis. Most of the common clinically significant hemoglobin (Hb) variants (ie, HbS, HbC, HbE, and others) are easily distinguished by hemoglobin electrophoresis and do not require molecular analysis. In addition, they are frequently found in complex hemoglobin disorders due to multiple genetic variants, and accurate classification requires sequencing data within the context of protein data. In some instances, beta-globin sequencing is necessary to identify or confirm the identity of rare variants, especially those associated with erythrocytosis and chronic hemolytic anemia. Rare hyper-unstable variants (also termed dominant beta thalassemia mutations) result in hemolytic anemia and do not create protein stable enough to be detectable by protein methods, including stability studies. They are not always associated with elevated HbA2 or microcytosis and, therefore, can be electrophoretically silent. These require a high degree of clinical suspicion as all electrophoretic testing as well as stability studies cannot exclude this condition.

Beta thalassemia is an autosomal recessive condition characterized by decreased or absent synthesis of beta-globin chains due to alterations in the beta-globin gene (*HBB*). No abnormal protein is present and diagnosis by electrophoresis relies on hemoglobin fraction percentage alterations (ie, HbA2 or HbF elevations).

Beta-thalassemia can be split into 3 broad classes (categorized by clinical features):

1. Beta thalassemia trait (also called beta thalassemia minor and beta thalassemia carrier) (B[A]B[+] or B[A]B[0]).
2. Beta thalassemia intermedia (B[+]B[+] or B[+]B[0])
3. Beta thalassemia major (B[+]B[0] or B[0]B[0])

Beta thalassemia trait is typically a harmless condition with varying degrees of microcytosis and hypochromia and sometimes mild anemia. Transfusions are not required. Beta thalassemia intermedia is a clinical distinction and is characterized by a more severe degree of anemia than beta thalassemia trait with few or intermittent transfusions required. Later in life, these individuals are at risk for iron overload even in the absence of chronic transfusion due to increased intestinal absorption of iron. Beta thalassemia major typically comes to medical attention early in life due to severe anemia, hepatosplenomegaly, and failure to thrive. Skeletal changes are also common due to expansion of the bone marrow. Without appropriate treatment these patients have a shortened lifespan.

The majority of beta-thalassemia variations (>90%) are point alterations, small deletions, or insertions, which are detected by beta-globin gene sequencing. The remaining beta-thalassemia sequence variants are either due to large genomic deletions of *HBB* or, very rarely, trans-acting beta thalassemia variations located outside of the beta-globin gene cluster. Some rare beta-chain variants can be clinically or electrophoretically indistinguishable from beta thalassemia and cannot be confirmed without molecular analysis.

Reference Values

Only orderable as a reflex. For more information see:

- HAEV1 / Hemolytic Anemia Evaluation, Blood
- HBEL1 / Hemoglobin Electrophoresis Evaluation, Blood
- MEV1 / Methemoglobinemia Evaluation, Blood
- REVE2 / Erythrocytosis Evaluation, Blood
- THEV1 / Thalassemia and Hemoglobinopathy Evaluation, Blood and Serum

An interpretive report will be provided.

Interpretation

The alteration will be provided with the classification, if known. Further interpretation requires correlation with protein studies and red blood cell indices.

Cautions

This assay will not detect large deletions or duplications within the beta-globin gene. In addition, hybrid beta-globin variants (ie, Hb Lepore) will not be detected by this method. This method cannot distinguish between homozygous and compound heterozygous variants associated with large deletions. This method cannot distinguish between double substitution on single chromosome and a compound heterozygous state. Beta-globin sequencing alone is not able to distinguish between alterations that are found in the same copy of the *HBB* gene (ie, variants that are "linked" or "in cis") and alterations found on different *HBB* gene copies (ie, are "in trans"). This limitation of sequencing may complicate diagnosis and has implications for inheritance and proper genetic counseling. To resolve these cases, molecular results must be correlated with electrophoretic and protein data, other laboratory findings, clinical findings, and family studies. Misinterpretation of results may occur if the information provided is inaccurate or incomplete.

Rare genetic alterations exist that could lead to false-negative or false-positive results. If results obtained do not match the clinical findings, additional testing should be considered.

Clinical Reference

1. Hoyer JD, Hoffman DR: The thalassemia and hemoglobinopathy syndromes. In: McClatchey KD, ed. Clinical Laboratory Medicine. 2nd ed. Lippincott Williams and Wilkins; 2002;866-895
2. Thein SL: The molecular basis of beta-thalassemia. Cold Spring Harb Perspect Med. 2013 May 1;1;3(5):a011700
3. Hoyer JD, Kroft, SH: Color Atlas of Hemoglobin Disorders: A Compendium Based on Proficiency Testing. CAP; 2003
4. Merchant S, Oliveira JL, Hoyer JD, Viswanatha DS: Molecular diagnosis in hematopathology. In: Goldblum J, His E, eds. Hematopathology: A Volume in Foundations in Diagnostic Pathology Series. 2nd ed. Churchill Livingstone; 2012

Performance

Method Description

Genomic DNA is extracted from whole blood. The *HBB* gene is amplified by polymerase chain reaction (PCR). The PCR product is then purified and sequenced in both directions using fluorescent dye-terminator chemistry. Sequencing products are separated on an automated sequencer and trace files analyzed for variations in all exons, introns with the exception of IVS-II-82 through IVS-II-650, the 5'UTR, the 3'UTR, and the promoter region. Results are correlated with routine studies to identify unusual beta globin variants.(Reddy PL, Bowie LJ: Sequence-based diagnosis of hemoglobinopathies in the clinical laboratory. Clin Lab Med. 1997;17[1]:85-96; Traeger-Synodinos J, Hartevelde CL: Advances in technologies for screening and diagnosis of hemoglobinopathies. Biomarkers Med. 2014;8[1]:115-127)

PDF Report

No

Day(s) Performed

Monday through Friday

Report Available

10 days

Specimen Retention Time

Blood: 2 weeks; DNA: 3 months

Performing Laboratory Location

Mayo Clinic Laboratories - Rochester Main Campus

Fees & Codes

Fees

- Authorized users can sign in to [Test Prices](#) for detailed fee information.
- Clients without access to Test Prices can contact [Customer Service](#) 24 hours a day, seven days a week.
- Prospective clients should contact their account representative. For assistance, contact [Customer Service](#).

Test Classification

This test was developed and its performance characteristics determined by Mayo Clinic in a manner consistent with CLIA requirements. It has not been cleared or approved by the US Food and Drug Administration.

CPT Code Information

81364-HBB (hemoglobin, beta) full sequence

LOINC® Information

Test ID	Test Order Name	Order LOINC® Value
WBSQR	Beta Globin Gene Sequencing, B	50996-8

Result ID	Test Result Name	Result LOINC® Value
47954	Beta Globin Gene Sequencing Result	50397-9
47955	Interpretation	59466-3