

## Overview

### Useful For

Management and triage of patients age 21 years or older with abnormal Papanicolaou (Pap) results

Diagnostic test for detection of human papillomavirus (HPV) high-risk genotypes associated with the development of cervical cancer

Results can be used as an aid in triaging women with abnormal Pap smear results

Individual genotyping of HPV-16 or HPV-18, if present

Results of HPV-16 and HPV-18 genotyping can be used as an aid in triaging women with positive high-risk HPV (HR-HPV) but negative Pap smear results

### Reflex Tests

Test ID	Reporting Name	Available Separately	Always Performed
TPDPC	Physician Interp Diagnostic	No	No
HPV	HPV with Genotyping, PCR, ThinPrep	Yes	No

### Testing Algorithm

When this test is ordered, a ThinPrep Pap screen will be performed. If the results include the criteria below, a high-risk human papillomavirus test will be performed:

- Atypical cells of undetermined significance (ASCUS), and the patient is 21 years or older
- Atypical squamous cells, cannot exclude high-grade squamous intraepithelial lesion (ASC-H) and the patient is 21 years or older
- Low-grade squamous intraepithelial lesion (LSIL), and the patient is 50 years or older
- Inadequate endocervical/transformation zone component, negative for intraepithelial lesion or malignancy, and the patient is 30 years or older

If ThinPrep Pap results are abnormal, a pathologist will review the case at an additional charge.

### Special Instructions

- [Gyn-Cytology Patient Information](#)

### Method Name

ThinPrep Pap Cytology Screening by Light Microscopy with HPV High-Risk DNA Detection with Genotyping by Real-Time Polymerase Chain Reaction

### NY State Available

Yes

## Specimen

### Specimen Type

Varies

### Advisory Information

Mayo Clinic Laboratories' clients need prior laboratory approval to order cytology testing.

### Necessary Information

1. An acceptable cytology request form must accompany specimen containers and include the following: Patient's name, medical record number, date of birth, sex, source (exact location and procedure used), date specimen was taken, name of ordering physician and pager number.

2. Submit any pertinent history or clinical information.

### Specimen Required

**Patient Preparation:** For optimal interpretation, Papanicolaou smears should be collected near the middle of the menstrual cycle. No douching, lubricant use, and sexual intercourse for 24 hours prior to specimen collection.

**Only 1 aliquot may be removed from PreservCyt sample vial prior to performing the ThinPrep Pap Test, regardless of the volume of the aliquot (maximum aliquot volume=4 mL).**

**Submit only 1 of the following specimens:**

**Specimen Type:** Cervical

**Supplies:** Thin Prep Media with Broom Kit (T056)

**Collection Container/Tube:** ThinPrep

**Specimen Volume:** 16 mL

### Collection Instructions:

1. Obtain adequate sampling from cervix using a broom-like collection device. If desired, use lukewarm water to warm and lubricate the speculum. Insert the central bristles of the broom into the endocervical canal deep enough to allow the shorter bristles to fully contact the ectocervix. Push gently, and rotate the broom in a clockwise direction 5 times.
2. Rinse the broom as quickly as possible into the PreservCyt solution vial by pushing broom into bottom of vial 10 times, forcing the bristles apart.
3. As a final step, swirl broom vigorously to further release material. Discard the collection device.
4. Tighten cap on vial so that the torque line on the cap passes the torque line on the vial.
5. **Specimen vial must be labeled with a minimum of 2 unique identifiers** (patient's name and medical record number or date of birth).

**Specimen Type:** Ectocervix and endocervix

**Supplies:** Thin Prep Media with Spatula and Brush Kit (T434)

**Collection Container/Tube:** ThinPrep

**Specimen Volume:** 16 mL

**Collection Instructions:**

1. Obtain an adequate sampling from the ectocervix using a plastic spatula. If desired, use lukewarm water to warm and lubricate the speculum. Select contoured end of plastic spatula and rotate it 360 degrees around the entire exocervix while maintaining tight contact with exocervical surface.
2. Rinse spatulas quickly as possible into the PreservCyt solution vial by swirling spatula vigorously in vial 10 times. Discard the spatula.
3. Next, obtain an adequate specimen from endocervix using an endocervical brush device. Insert the brush into the cervix until only the bottommost fibers are exposed. Slowly rotate one-quarter or one-half turn in 1 direction. **Do not overrotate.**
4. Rinse the brush as quickly as possible in the PreservCyt solution by rotating the device in the solution 10 times while pushing against the PreservCyt vial wall.
5. Swirl brush vigorously as final step to further release material. Discard the brush.
6. Tighten the cap so that the torque line on the cap passes the torque line on the vial.
7. **Specimen vial must be labeled with a minimum of 2 unique identifiers** (patient's name and medical record number or date of birth).

**Forms**

[Gyn-Cytology Patient Information](#) (T601) in Special Instructions

**Specimen Minimum Volume**

See Specimen Required

**Reject Due To**

Other	SurePath vials
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**Specimen Stability Information**

Specimen Type	Temperature	Time	Special Container
Varies	Ambient (preferred)	42 days	THIN PREP
	Refrigerated	42 days	THIN PREP

**Clinical and Interpretive**

**Clinical Information**

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Squamous cell carcinoma of the cervix is believed to develop in progressive stages from normal through precancerous (dysplastic) stages, to carcinoma in situ, and eventually invasive carcinoma. This sequence is felt to develop over a matter of years in most patients.

Follow-up of the cervical Pap abnormality atypical squamous cells of undetermined significance (ASCUS) is costly and frustrating to patients and clinicians because a large percentage of these patients have normal colposcopic and biopsy findings. Yet, a significant percentage (10%-15%) will have an underlying high-grade squamous intraepithelial lesion (HSIL).

The majority (>99%) of cervical epithelial neoplasms are the result of human papillomavirus (HPV) infection. High-risk HPV (HR-HPV) types (16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, and 68) can result in both low-grade squamous intraepithelial lesions and HSIL, as well as invasive carcinomas. Patients with HSIL have a greater risk for progression to carcinoma.

In the setting of an abnormal Pap result, the presence of HR-HPV types in cervical specimens identifies a subgroup of patients with a greater likelihood of having a HSIL.

If the patient has been previously diagnosed with an abnormal Pap result or is at high risk, considering ordering this test, which is diagnostic, rather than the screen (STHPV / ThinPrep Screen with Human Papillomavirus [HPV] Reflex).

Persistent infection with HPV is the principal cause of cervical cancer and its precursor cervical intraepithelial neoplasia (CIN).(1-3) The presence of HPV has been implicated in more than 99% of cervical cancers worldwide. HPV is a small, nonenveloped, double-stranded DNA virus, with a genome of approximately 8,000 nucleotides. There are more than 118 different types of HPV and approximately 40 different HPVs that can infect the human anogenital mucosa. However, data suggest that 14 of these types (HPV types 16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 66, and 68) are considered high-risk for the development of cervical cancer and its precursor lesions. Furthermore, HPV types 16 and 18 have been regarded as the genotypes most closely associated with progression to cervical cancer. HPV-16 is the most carcinogenic, and is associated with approximately 60% of all cervical cancers, while HPV-18 accounts for approximately 10% to 15% of cervical cancers.(4-6)

Although persistent infection with HR-HPV is necessary for the development of cervical cancer and its precursor lesions, only a very small percentage of infections progress to these disease states. Sexually transmitted infection with HPV is extremely common, with estimates of up to 75% of all women being exposed to HPV at some point. However, almost all infected women will mount an effective immune response and clear the infection within 2 years without any long-term health consequences. An infection with any HPV type can produce CIN although this also usually resolves once the HPV infection has been cleared.

In developed countries with cervical cancer screening programs, the Pap smear has been used since the mid-1950s as the primary tool to detect early precursors to cervical cancer. Although it has decreased the death rates due to cervical cancer dramatically in those countries, the Pap smear and subsequent liquid-based cytology methods require subjective interpretation by highly trained cytopathologists and misinterpretation can occur. Cytological abnormalities are primarily due to infection with HPV; however, various inflammatory conditions or sampling variations can result in false-positive cytology results. Triage of an abnormal cytology result may involve repeat testing, colposcopy and biopsy. A histologically confirmed high-grade lesion must be surgically removed or ablated in order to prevent the development of invasive cervical cancer.

Nucleic acid (DNA) testing by PCR has become a standard, noninvasive method for determining the presence of a cervical HPV infection. Proper implementation of nucleic acid testing for HPV may 1) increase the sensitivity of cervical cancer screening programs by detecting high-risk lesions earlier in women 30 years and older with normal cytology and 2) reduce the need for unnecessary colposcopy and treatment in patients 21 and older with cytology results showing atypical squamous cells of undetermined significance (ASC-US).

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Recently, data suggest that individual genotyping for HPV types 16 and 18 can assist in determining appropriate follow-up testing and triaging women at risk for progression to cervical cancer. Studies have shown that the absolute risk of CIN-2 or worse in HPV-16 or HPV-18 positive women is 11.4% (95% confidence interval [CI] 8.4%-14.8%) compared with 6.1% (95% CI, 4.9%-7.2%) of women positive for other HR-HPV genotypes and 0.8% (95% CI, 0.3%-1.5%) in HR-HPV negative women.(7) Based in part on these data, the American Society for Colposcopy and Cervical Pathology (ASCCP) now recommends that HPV 16 and 18 genotyping be performed on women who are positive for HR-HPV but negative by routine cytology. Women who are found to be positive for HPV-16 or HPV-18 may be referred to colposcopy, while women who are negative for genotypes 16 and 18 may have repeat cytology and HR-HPV testing in 12 months.(4)

## Reference Values

ThinPrep PAPANICOLAOU

Satisfactory for evaluation. Negative for intraepithelial lesion or malignancy.

**Note:** Abnormal results will be reviewed by a pathologist at an additional charge.

HUMAN PAPILLOMAVIRUS (HPV)

Negative for HPV genotypes 16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 66, and 68

## Interpretation

Cytology:

Standard reporting, as defined by the Bethesda System (TBS) is utilized.(8)

Human papillomavirus (HPV):

A positive result indicates the presence of HPV DNA due to 1 or more of the following genotypes: 16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 66, and 68.

A negative result indicates the absence of HPV DNA of the targeted genotypes.

For patients with atypical squamous cells of undetermined significance (ASC-US) Papanicolaou (Pap) smear result and who are positive for high-risk HPV (HR-HPV), consider referral for colposcopy, if clinically indicated.

For women aged 30 years and older with a negative Pap smear result, but who are positive for HPV-16 or HPV-18, consider referral for colposcopy, if clinically indicated.

For women aged 30 years and older with a negative Pap smear, positive HR-HPV test result, but who are negative for HPV-16 and HPV-18, consider repeat testing by both cytology and a HR-HPV test in 12 months.

## Cautions

The Papanicolaou (Pap) test is a screening test for cervical cancer with inherent false-negative results. A negative human papillomavirus (HPV) test or Pap smear result does not preclude the presence of carcinoma or intraepithelial lesion. The false-negative rates of the Pap test range from 15% to 30%.

The cobas HPV test detects DNA of the high-risk types 16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 66, and 68. This test does not detect DNA of HPV low-risk types (eg, 6, 11, 42, 43, 44) since these are not associated with cervical cancer and its precursor lesions.

The cobas HPV test is not recommended for evaluation of suspected sexual abuse.

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Prevalence of HPV infection in a population may affect performance. Positive predictive values decrease when testing populations with low prevalence or individuals with no risk of infection.

Infection with HPV is not an indicator of cytologic high-grade intraepithelial lesion (HSIL) or underlying high-grade cervical intraepithelial neoplasia (CIN), nor does it imply that CIN2-3 or cancer will develop. Most women infected with 1 or more high-risk HPV (HR-HPV) types do not develop CIN2-3 or cancer.

A negative HR-HPV result does not exclude the possibility of future cytologic HSIL or underlying CIN2-3 or cancer.

### **Clinical Reference**

1. Solomon D, Schiffman M, Tarone R: Comparison of three management strategies for patients with atypical squamous cells of undetermined significance: baseline results from a randomized trial. *J Natl Cancer Inst* 2001;93:293-299
2. Soloman D, Davey D, Kurman R, et al: The 2001 Bethesda System: terminology for reporting results of cervical cytology. *JAMA* 2002;287:2114-2119
3. Wright TC, Cox JT, Massad LS, et al: 2001 Consensus Guidelines for the management of women with cervical cytological abnormalities. *JAMA* 2002;287:2120-2129
4. Saslow D, Solomon D, Lawson HW, et al: American Cancer Society, American Society for Colposcopy and Cervical Pathology, and American Society for Clinical Pathology Screening Guidelines for the Prevention and Early Detection of Cervical Cancer. *J Low Genit Tract Dis* 2012;16(3):175-204
5. Walboomers JM, Jacobs MV, Manos MM, et al: Human papillomavirus is a necessary cause of invasive cervical cancer worldwide. *J Pathol* 1999;189:12-19
6. de Sanjose S, Quint WG, Alemany L, et al: Human papillomavirus genotype attribution in invasive cervical cancer: a retrospective cross-sectional worldwide study. *Lancet Oncol* 2010;11:1048-1056
7. Wright TC, Stoler MH, Sharma A, et al: Evaluation of HPV-16 and HPV-18 genotyping for the triage of women with high-risk HPV positive, cytology-negative results. *Am J Clin Pathol* 2011 Oct;136(4):578-586
8. Solomon D: Foreword; in Nayar R, Wilbur DC (eds): *The Bethesda System for Reporting Cervical Cytology: Definitions, Criteria, and Explanatory Notes, Third Edition*. New York, Springer 2015.
9. Massad LS, Einstein MH, Huh WK, et al: 2012 updated consensus guidelines for the management of abnormal cervical cancer screening tests and cancer precursors. *J Low Genit Tract Dis* 2013 April;17(5 Suppl 1):S1-S27
10. Sherman ME, Lorincz A, Scott DR, et al: Baseline cytology, human papillomavirus testing, and risk for cervical neoplasia: a 10-year cohort analysis. *J Nat Cancer Inst* 2003 January;95(1):46-52

### **Performance**

#### **Method Description**

A ThinPrep Pap specimen is collected, processed on a T2000 or T5000 processor, and stained with a Pap stain. Cases are examined microscopically and those with appropriate cytologic diagnoses are referred for human papillomavirus (HPV) testing if the patient is 21 or older. (Operator's manual: ThinPrep 2000 System, Cytyc, Marlboro, MA; Operator's Manual: ThinPrep 5000 Processor, Cytyc, Marlboro, MA)

The Cobas human papillomavirus (HPV) test targets and detects nucleic acid from the L1 region of the HPV genome

using real-time polymerase chain reaction (PCR) technology. The Cobas HPV test is used for the in vitro qualitative detection of 14 high-risk HPV types commonly associated with cervical cancer. The assay is able to specifically assess for the presence or absence of HPV genotypes 16 and 18, while concurrently detecting the remaining 12 high-risk types (31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 66 and 68). The Cobas HPV test is used in conjunction with the Cobas 4800 System. The Cobas 4800 System comprises the Cobas x 480 instrument and Cobas z 480 analyzer that fully automates the Cobas HPV from sample extraction through amplification, detection, and data reduction. (Procedure manual and package insert: Cobas HPV test. Roche Diagnostics. Indianapolis, IN, version 05641268001-01EN)

**PDF Report**

No

**Day(s) and Time(s) Test Performed**

Monday through Friday; Varies

**Analytic Time**

5 days

**Maximum Laboratory Time**

8 days

**Specimen Retention Time**

14 days after report issued if HPV testing has not been performed. 1 week if HPV testing has been performed.

**Performing Laboratory Location**

Rochester

**Fees and Codes****Fees**

- Authorized users can sign in to [Test Prices](#) for detailed fee information.
- Clients without access to Test Prices can contact [Customer Service](#) 24 hours a day, seven days a week.
- Prospective clients should contact their Regional Manager. For assistance, contact [Customer Service](#).

**Test Classification**

This test has been cleared or approved by the U.S. Food and Drug Administration and is used per manufacturer's instructions. Performance characteristics were verified by Mayo Clinic in a manner consistent with CLIA requirements.

**CPT Code Information**

88142

88141-TPDPC (if appropriate)

**LOINC® Information**

Test ID	Test Order Name	Order LOINC Value
DTHPV	ThinPrep Diagnostic HPV Reflex	In Process



Result ID	Test Result Name	Result LOINC Value
71321	Interpretation	59465-5
71322	Participated in the Interpretation	No LOINC Needed
71323	Report electronically signed by	19139-5
71324	Addendum	35265-8
71325	Gross Description	22634-0
CY012	Pap Test Source	19763-2
CY013	Clinical History	22636-5
CY014	Menstrual Status(LMP, PM, Pregnant)	8678-5
CY015	Hormone Therapy/Contraceptives	8659-5
71577	Disclaimer	62364-5
71823	Case Number	80398-1