Overview

Useful For
Aids in the diagnosis of amebic keratitis in conjunction with clinical findings

Highlights
This assay is intended as an aid in the diagnosis of amebic keratitis (AK) in conjunction with clinical findings.

This test has similar sensitivity and specificity to culture but provides a more rapid result.

Acanthamoeba species are free-living organisms and may be found widely in the environment.

Method Name
Real-Time Polymerase Chain Reaction (PCR)/TaqMan DNA Probe Hybridization

NY State Available
Yes

Specimen

Specimen Type
Varies

Advisory Information
Although verification experiments did not detect Acanthamoeba species DNA in contact lenses from asymptomatic adults, it is possible that PCR may detect asymptomatic colonization/contamination and, therefore, testing should not be performed on asymptomatic individuals.

Necessary Information
Specimen source is required.

Specimen Required
The preferred specimen for Acanthamoeba PCR from an ocular source is corneal scraping or biopsy.

Submit only 1 of the following specimens:

Specimen Type: Tissue, fresh
Sources: Ocular
Container/Tube: Sterile container
Specimen Volume: 5-10 mm

Collection Instructions: Submit tissue in a sterile container with 1 mL of sterile saline, minimal essential media (MEM), or viral transport media.

Preferred Paraffin-embedded tissue block:
Test Definition: ACARP  
Acanthamoeba species Detection, PCR

**Supplies:** Tissue Block Container (T553)

**Specimen Type:** Formalin-fixed, paraffin-embedded tissue block (FFPE)

**Sources:** Ocular

**Container/Tube:** Tissue block

**Collection Instructions:** Submit a FFPE tissue block to be cut and returned.

**Specimen Stability Information:** Ambient (preferred)/Refrigerated

**Acceptable Paraffin-embedded tissue block:**

**Specimen Type:** Formalin-fixed, paraffin-embedded tissue block (FFPE)

**Sources:** Ocular

**Container/Tube:** Sterile container for each individual cut section (scroll).

**Collection Instructions:** Perform microtomy and prepare five separate 10-micron sections. Each section (scroll) must be placed in a separate sterile container for submission.

**Specimen Stability Information:** Ambient (preferred)/Refrigerated

**Specimen Type:** Scrapings, swabs

**Sources:** Eye, ocular, cornea

**Container/Tube:** Sterile container

**Specimen Volume:** 1 mL

**Collection Instructions:**

1. Collect corneal scrapings using a scalpel or other sharp device to remove the outer layer of cells from the eye.
2. Swish the collection device in 1 mL of sterile saline, minimal essential media (MEM), or viral transport media.
3. Remove the collection device from the collection container before submitting to the lab.
4. **Specimens containing scalpel blades will be canceled.**

**Additional Information:** Swabs are not the preferred specimen for this test and may yield false-negative results. **Specimens collected using wooden shafted swabs and calcium alginate-tipped swabs will be canceled.**

**Specimen Type:** Contact lenses

**Container/Tube:** Sterile container
**Specimen Volume:** Entire collection

**Collection Instructions:**

1. Place entire contact lens in a sterile container with 1 mL sterile saline, contact lens solution, viral transport media, or minimal essential media (MEM).

2. Right and Left lenses must be submitted individually using multiple sterile containers or in the original contact lens case. **Multiple orders must be created.**

3. Indicate Right or Left in the specimen source.

**Specimen Type:** Contact lens solution

**Container/Tube:** Sterile container

**Specimen Volume:** 1 mL solution

**Specimen Type:** Contact lens cases without lenses

**Container/Tube:** Sterile container

**Specimen Volume:** 1 mL solution or entire case

**Additional Information:**

1. Depending on the type of case submitted, it may be necessary to test right and left chambers individually. **Multiple orders must be created.**

2. Indicate Right or Left in the specimen source.

**Forms**

If not ordering electronically, complete, print, and send a [Microbiology Test Request](#) (T244) with the specimen.

**Specimen Minimum Volume**

- Tissue: 5 mm biopsy
- Scrapings: 0.5 mL
- Contact Lens Solution: 1 mL

**Reject Due To**

| Other                                      | Calcium alginate-tipped swab, wood swab, or transport swab containing gel; specimens containing scalpel blades, unstained slides |

**Specimen Stability Information**

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<tr>
<th>Specimen Type</th>
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Clinical and Interpretive

Clinical Information

*Acanthamoeba* are ubiquitous, free-living, microscopic amebae that cause rare, but severe, infections of the eye, skin, lungs, and central nervous system (CNS). They are found worldwide in water and soil and may enter the body through inhalation, contamination of wounds, and contact lens use. As many as 24 species comprising 18 genotypes (T1-T18) have been described, although most human infections are due to genotype T4. Given their widespread distribution in the environment, many people will be exposed to *Acanthamoeba* during their lifetime, but very few will become sick from this exposure.

The most common form of *Acanthamoeba* infection is amebic keratitis (AK). Infection occurs primarily in contact lens wearers due to contamination of lenses, cleaning solutions, or storage cases. Amebae can also enter the cornea following trauma. AK is a painful, subacute corneal infection associated with extensive scarring and blindness if untreated. Cases generally respond to treatment but relapse is common. Compared to corneal infection, involvement of the CNS is rare and seen primarily in severely immunocompromised individuals such as organ transplant recipients and patients with AIDS. CNS infection may also be caused by a related ameba, *Balamuthia mandrillaris*.

AK is usually clinically suspected based on symptoms and confocal ophthalmologic examination. Confirmation of infection is classically identified by microscopic examination and culture of corneal tissue and contact lenses or equipment using tap water agar plate overlain with bacteria as a food source for the amebae. Unfortunately, it must be held and examined for 7 days for maximum sensitivity. PCR provides a more rapid result with similar sensitivity to culture and is, therefore, the preferred method for confirming the clinical diagnosis of AK.

Reference Values

Negative

Interpretation

A positive result indicates the presence of *Acanthamoeba* species DNA and is consistent with active or recent infection. While positive results are highly specific indicators of disease, they should be correlated with symptoms, clinical findings, and confocal ophthalmologic examination.

Cautions

While this assay is designed to detect symptomatic infection with *Acanthamoeba* species, the widespread distribution of these free-living microscopic amebae in the environment may contaminate inanimate objects such as contact lenses and cases. Thus, it should be used for patients with a clinical history and ocular symptoms consistent with amebic keratitis.

Inadequate specimen collection or improper storage may invalidate test results.

*Acanthamoeba* species DNA may be detectable for an unknown period of time after adequate treatment.

Supportive Data

The following assay verification data supports the use of this assay for clinical testing.

Species Inclusivity:

The *Acanthamoeba* PCR assay detects the 20 different strains of *Acanthamoeba*, including the genotypes that cause human disease.

Accuracy/Diagnostic Sensitivity and Specificity-Fresh Specimens:
Results from this PCR assay detecting the 18S rRNA gene of *Acanthamoeba* species were compared to culture results on 112 contact/ocular specimens. Of the 12 specimens that were positive by culture, 11 were detected by PCR (sensitivity 92%). PCR also detected an additional 2 positive specimens, which were both from the same patient with a clinical diagnosis of amebic keratitis (AK), while 98 specimens were negative by both culture and PCR (specificity 98%).

Accuracy/Diagnostic Sensitivity and Specificity-Formalin-Fixed Paraffin-Embedded (FFPE) Specimens:

Twenty-four FFPE archived tissue blocks were tested by the *Acanthamoeba* species PCR assay and results were compared to histopathologic (light microscopic) diagnosis. Fourteen of the tissues had a morphologic diagnosis of acanthamebic keratitis; of these, 12 were positive by PCR (sensitivity 86%). Ten specimens were negative by both histopathology and PCR (specificity 100%).

Supplemental Accuracy Data:

Spiking studies were performed using ocular material in transport media (contact lens fluid, MEM), fresh tissue, and FFPE tissue spiked with *Acanthamoeba* genomic DNA at an approximate concentration of 50 targets/mcL. All samples were then extracted and tested in a blinded fashion. At 50 targets/mcL, 100% of the ocular material, the fresh, and the FFPE tissue were positive by PCR.

Analytical Sensitivity/Limit of Detection (LoD):

- The LoD determined with serial dilution of cultured *Acanthamoeba* cysts (counted using a hemocytometer) was considered to be 1 cyst per processed sample.

- The LoD established using genomic DNA spiked into contact lens solution/MEM transport media is 1.26 target copies/mcL.

- The LoD established using genomic DNA spiked into fresh tissue matrix is 6.5 target copies/mL.

- The LoD established using genomic DNA spiked into FFPE tissue matrix is 5.7 target copies/mL

Analytical Specificity:

No PCR signal was obtained from the extracts of 47 bacterial, viral, parasitic, and fungal isolates from similar organisms or from organisms commonly found in the specimens tested.

Precision:

Qualitative inter- and intra-assay precision was 100%. All crossing point values were within 2 cycles of the mean.

Reference Range:

The reference range is "Negative" for this assay. PCR and culture performed on 291 contact lenses from asymptomatic individuals failed to detect *Acanthamoeba* DNA or growth.

However, PCR may detect *Acanthamoeba* species colonization due to the widespread distribution of this free-living ameba in the environment, and PCR testing should only be used for patients with a clinical history and ocular symptoms consistent with AK.

Reportable Range:
This is a qualitative assay, and the results are reported as negative or positive for targeted *Acanthamoeba* species.
Clinical Reference


Performance

Method Description

The assay is performed on the Roche LightCycler (LC) 480 II instrument, following DNA extraction on the Roche MagNA Pure or the Siemens Tissue Preparation System (TPS). The LC 480 II instrument is an automated instrument that amplifies and monitors the development of target nucleic acid (amplicon) after each cycle of PCR.

The DNA target for this PCR assay is a gene encoding the nuclear small subunit ribosomal 18S rRNA.

The PCR mix contains a forward and reverse primer specific for Acanthamoeba species template amplification and 1 TaqMan probe (CY5). The CY5 probe contains a fluorophore (5’-end) and a quencher (3’-end) in close proximity; the quencher inhibits the fluorescence signal from the fluorophore while the probe is intact. After the probe anneals to the targeted Acanthamoeba 18S rRNA, it is subsequently degraded by a DNA polymerase with 5’-3’ exonuclease activity, resulting in release of the fluorophore and production of detectable fluorescent signal.(Qvarnstrom Y, Visvesvara GS, Sriram R, da Silva AJ: Multiplex real-time PCR assay for simultaneous detection of Acanthamoeba spp, Balamuthia mandrillaris, and Naegleria fowleri. J Clin Microbiol 2006;44:3589-3595)

PDF Report

No

Day(s) and Time(s) Test Performed

Monday through Saturday

Analytic Time

2 days

Maximum Laboratory Time

3 days

Specimen Retention Time

7 Days

Performing Laboratory Location

Rochester

Fees and Codes

Fees

- Authorized users can sign in to Test Prices for detailed fee information.
- Clients without access to Test Prices can contact Customer Service 24 hours a day, seven days a week.
- Prospective clients should contact their Regional Manager. For assistance, contact Customer Service.
Test Definition: ACARP
Acanthamoeba species Detection, PCR

Test Classification
This test was developed and its performance characteristics determined by Mayo Clinic in a manner consistent with CLIA requirements. This test has not been cleared or approved by the U.S. Food and Drug Administration.

CPT Code Information
87798

LOINC® Information

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