Overview

Useful For

Detection of *Chlamydia trachomatis* or *Neisseria gonorrhoeae*

Profile Information

<table>
<thead>
<tr>
<th>Test ID</th>
<th>Reporting Name</th>
<th>Available Separately</th>
<th>Always Performed</th>
</tr>
</thead>
<tbody>
<tr>
<td>CTRNA</td>
<td>Chlamydia trachomatis Amplified RNA</td>
<td>Yes</td>
<td>Yes</td>
</tr>
<tr>
<td>GCRNA</td>
<td>Neisseria gonorrhoeae Amplified RNA</td>
<td>Yes</td>
<td>Yes</td>
</tr>
</tbody>
</table>

Method Name

Transcription Mediated Amplification

NY State Available

Yes

Specimen

Specimen Type

Varies

Necessary Information

Specimen source is required.

Specimen Required

Submit only 1 of the following specimens:

**Swab specimen must be collected** using an Aptima Collection Unisex Swab (T583) or Aptima Collection Multitest Swab (T584, formerly called Aptima Vaginal Swab Specimen Collection Kit). These swabs are contained in the Aptima Collection Kit.

**Supplies:** Swab, Aptima Male/Female Collection (T583)

**Specimen Type:** Endocervix

**Container/Tube:** Aptima Collection Unisex Swab (T583)

**Specimen Volume:** Swab

**Collection Instructions:**

1. Use cleaning swab (white shaft) to remove excess mucus from endocervix and discard.
2. Insert second swab (blue shaft) 1 to 1.5 cm into endocervical canal, and rotate swab gently for 30 seconds. Avoid touching vaginal wall when removing swab.

3. Place second swab into transport tube provided in collection kit. Snap off swab at score line so swab fits into closed tube.

4. Cap tube securely, and label tube with patient's entire name, and date and time of collection.

5. Transport and store swab container at 2 to 30 degrees C (refrigerate is preferred temperature) within 60 days of collection. If longer storage is needed, freeze at -20 to -70 degrees C for 12 months.

**Supplies:** Swab, Aptima Multitest Swab Specimen Collection Kit (T584)

**Specimen Type:** Vaginal

**Container/Tube:** Aptima Collection Multitest Swab (T584)

**Specimen Volume:** Swab

**Collection Instructions:**

1. Insert swab (pink shaft) about 5 cm past introitus and rotate gently for 30 seconds.

2. Place swab into transport tube provided in collection kit. Snap off swab at score line so swab fits into closed tube.

3. Cap tube securely, and label tube with patient's entire name, and date and time of collection.

4. Transport and store swab container at 2 to 30 degrees C (refrigerate is preferred temperature) within 60 days of collection. If longer storage is needed, freeze at -20 to -70 degrees C for 12 months.

**Supplies:** Swab, Aptima Male/Female Collection (T583)

**Specimen Type:** Urethra (Males Only)

**Container/Tube:** Aptima Collection Unisex Swab (T583)

**Specimen Volume:** Swab

**Collection Instructions:**

1. Patient should not have urinated for at least 1 hour prior to collection.

2. With a rotating movement, insert swab (blue shaft) 2 to 4 cm into urethra.

3. Once inserted, rotate swab gently at least 1 full rotation using sufficient pressure to ensure swab comes into contact with all urethral surfaces. Allow swab to remain inserted for 2 to 3 seconds.

4. Place swab in transport tube provided in collection kit. Snap off swab at score line so swab fits into closed tube.

5. Cap tube securely, and label tube with patient's entire name, and date and time of collection.
6. Transport and store swab container at 2 to 30 degrees C (refrigerate is preferred temperature) within 60 days of collection. If longer storage is needed, freeze at -20 to -70 degrees C for 12 months.

**Supplies:** Aptima Urine Transport Tube (T582)

**Specimen Type:** Urine (Males and Females)

**Container/Tube:** Aptima Urine Specimen Transport Tube (T582)

**Specimen Volume:** 15-20 mL

**Collection Instructions:**

1. Patient should not have urinated for at least 1 hour prior to specimen collection.

2. Patient should collect first portion of random voided urine (first part of stream) into a sterile, plastic, preservative-free container.

3. Transfer 2 mL of urine into the urine specimen transport tube using the disposable pipette provided within 24 hours of collection. The correct volume of urine has been added when the fluid level is between the black fill lines on the urine transport tube. Place the labels on the transport tube so the black fill lines are still visible for volume confirmation at Mayo Clinic Laboratories.

4. Transport and store urine specimen transport container at 2 to 30 degrees C (refrigerate is preferred temperature) within 30 days of collection. If longer storage is needed, freeze at -20 to -70 degrees C for 12 months.

**Supplies:** Aptima Thin Prep Transport Tube (T652)

**Specimen Type:** ThinPrep Specimen (Endocervix)

**Container/Tube:** ThinPrep (also called PreservCyt) Collection Kit

**Specimen Volume:** 1 mL

**Collection Instructions:**

1. Aliquot ThinPrep specimen for *Chlamydia* and/or *Neisseria* testing before processing for Pap smear. For each specimen, use a new pair of clean gloves.

2. Vortex ThinPrep/PreservCyt vial 3 to 10 seconds. Within 1 minute of vortexing:

   a. Transfer 1 mL of specimen into the Aptima Specimen Transfer Tube (T652) using a disposable transfer pipette or a pipette tip containing a filter (aerosol barrier or hydrophobic plug).

   b. Process only 1 ThinPrep and transfer tube set at a time.

   c. Recap Aptima Specimen Transfer Tube tightly and gently invert 3 times to mix.

3. Label Aptima transfer tube with appropriate label.
4. Use remainder of ThinPrep specimen for Pap testing.

5. Transport and store specimen transfer container at 2 to 30 degrees C (refrigerate is preferred temperature) within 60 days of collection. If longer storage is needed, freeze at -20 to -70 degrees C for 12 months.

**Forms**
If not ordering electronically, complete, print, and send a [Microbiology Test Request](#) (T244) with the specimen.

**Specimen Minimum Volume**
Endocervical in PreservCyt: 1mL
Urine: 2 mL
Swabs (Endocervical, Urethral, Vaginal): Entire Collection

**Reject Due To**

| Other | Midstream urine specimen Overfilled or underfilled urine transport tubes Specimen collected into a SurePath device Transport tubes containing a cleaning swab or more than 1 swab |

**Specimen Stability Information**

<table>
<thead>
<tr>
<th>Specimen Type</th>
<th>Temperature</th>
<th>Time</th>
<th>Special Container</th>
</tr>
</thead>
<tbody>
<tr>
<td>Varies</td>
<td>Refrigerated (preferred)</td>
<td>APTIMA VIAL</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Ambient</td>
<td>APTIMA VIAL</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Frozen</td>
<td>APTIMA VIAL</td>
<td></td>
</tr>
</tbody>
</table>

**Clinical and Interpretive**

**Clinical Information**

Chlamydia is caused by the obligate intracellular bacterium *Chlamydia trachomatis* and is the most prevalent sexually transmitted bacterial infection (STI) in the United States.(1,2) In 2010, 1.3 million documented cases were reported to the CDC.(2) Given that 3 out of 4 infected women and 1 out of 2 infected men will be asymptomatic initially, the actual prevalence of disease is thought to be much greater than reported.(2) The organism causes genitourinary infections in women and men and may be associated with dysuria and vaginal, urethral, or rectal discharge. In women, complications include pelvic inflammatory disease, salpingitis, and infertility. Approximately 25% to 30% of women who develop acute salpingitis become infertile.(2) Complications among men are rare, but include epididymitis and sterility. Rarely, genital chlamydial infection can cause arthritis with associated skin lesions and ocular inflammation (Reiter syndrome). *C trachomatis* can be transmitted from the mother during delivery and is associated with conjunctivitis and pneumonia. Finally, *C trachomatis* may cause hepatitis and pharyngitis in adult.

Once detected, the infection is easily treated by a short course of antibiotic therapy.(2) Annual *Chlamydia* screening is now recommended for all sexually active women age 25 years and younger, and for older women with risk factors for infection, such as a new sex partner or multiple sex partners.(2) The CDC also recommends that all pregnant women be given a screening test for chlamydia infection.(2) Repeat testing for test-of-cure is not recommended after treatment with a standard treatment regimen unless patient compliance is in question, reinfection is suspected, or the patient's symptoms persist. Repeat testing of pregnant women, 3 weeks after completion of therapy, is also recommended to ensure therapeutic cure.(2)
Gonorrhea is caused by the bacterium Neisseria gonorrhoeae. It is also a very common STI, with 301,174 cases of gonorrhea reported to CDC in 2009.(1,2) Like Chlamydia, many infections in women are asymptomatic, and the true prevalence of gonorrhea is likely much higher than reported.(1,2) The organism causes genitourinary infections in women and men and may be associated with dysuria and vaginal, urethral, or rectal discharge. Complications include pelvic inflammatory disease in women and gonococcal epididymitis and prostatitis in men. Gonococcal bacteremia, pharyngitis, and arthritis may also occur. Infection in men is typically associated with symptoms that would prompt clinical evaluation. Given the risk for asymptomatic infection in women, screening is recommended for women at increased risk of infection (eg, women with previous gonorrhea or other STI, inconsistent condom use, new or multiple sex partners, and women in certain demographic groups such as those in communities with high STI prevalence.)(2) The CDC currently recommends dual antibiotic treatment due to emerging antimicrobial resistance.(2)

Culture was previously considered to be the gold standard test for diagnosis of C trachomatis and N gonorrhoeae infection. However, these organisms are labile in vitro, and precise specimen collection, transportation, and processing conditions are required to maintain organism viability, which is necessary for successful culturing. In comparison, nucleic acid amplification testing (NAAT) provides superior sensitivity and specificity and is now the recommended method for diagnosis in most cases.(2-5) Immunoassays and nonamplification DNA tests are also available for C trachomatis and N gonorrhoeae detection, but these methods are significantly less sensitive and less specific than NAATs.(2-5)

Improved screening rates and increased sensitivity of NAAT testing have resulted in an increased number of accurately diagnosed cases.(2-5) Improved detection rates result from both the increased performance of the assay and the patients' easy acceptance of urine testing. Early identification of infection enables sexual partners to seek testing and/or treatment as soon as possible and reduces the risk of disease spread. Prompt treatment reduces the risk of infertility in women.

Reference Values

Chlamydia trachomatis

Negative

Neisseria gonorrhoeae

Negative

Interpretation

A positive result indicates that rRNA of Chlamydia trachomatis or Neisseria gonorrhoeae is present in the specimen tested and strongly supports a diagnosis of chlamydial or gonorrheal infection.

A negative result indicates that rRNA for C trachomatis or N gonorrhoeae was not detected in the specimen.

The predictive value of an assay depends on the prevalence of the disease in any particular population. In settings with a high prevalence of sexually transmitted disease, positive assay results have a high likelihood of being true-positives. In settings with a low prevalence of sexually transmitted disease, or in any setting in which a patient's clinical signs and symptoms or risk factors are inconsistent with gonococcal or chlamydial urogenital infection, positive results should be carefully assessed and the patient retested by other methods (eg, culture for N gonorrhoeae), if appropriate.

A negative result does not exclude the possibility of infection. If clinical indications strongly suggest gonococcal or chlamydial infection, additional specimens should be collected for testing. A result of indeterminate indicates that a new specimen should be collected.
This test has not been shown to cross react with commensal (nonpathogenic) *Neisseria* species present in the oropharynx.

**Cautions**

Care must be taken to avoid cross-contamination during handling of PreservCyt solution liquid Pap specimens. If testing PreservCyt specimens processed with the ThinPrep 2000 processor, it is important to follow procedures to reduce the risk for cross-contamination during Pap processing such as bleaching of the PreservCyt filter cap and changing gloves between each sample. Refer to the ThinPrep 2000 Processor Operator's Manual and the Aptima specimen for more guidance.

The performance of endocervical, vaginal, and male urethral swab specimens, male and female urine specimens, and PreservCyt solution liquid Pap specimens has not been evaluated in adolescents younger than 16 years of age. The performance of vaginal swab specimens has not been evaluated in pregnant women.

This report is intended for use in clinical monitoring or management of patients; it is not intended for use in medico-legal applications.

Appropriate specimen collection and handling is necessary for optimal assay performance.

Results should be interpreted in conjunction with other laboratory and clinical information.

A negative test result does not exclude the possibility of infection. Improper specimen collection, concurrent antibiotic therapy, presence of inhibitors, or low numbers of organisms in the specimen (ie, below the sensitivity of the test) may cause false-negative test results.

In low-prevalence populations, positive results must be interpreted carefully as false-positive results may occur more frequently than true-positive results in this setting.

In general, this assay should not be used to assess therapeutic success or failure, since nucleic acids from these organisms may persist for 3 weeks or more following antimicrobial therapy.

The presence of mucous does not interfere with this assay. However, this test requires endocervical cells, and if excess mucous is not removed prior to collection, adequate numbers of these cells may not be obtained.

No interference is expected due to:

-Blood

-Lubricants and spermicides

The effects of use of tampons, douching, specimen types other than those listed in Specimen Required, and specimen collection variable have not been determined.

Testing of urine specimens with this method is not intended to replace cervical exam and endocervical sampling for diagnosis of urogenital infection; infections may result from other causes or concurrent infections may occur.

Testing urine specimens as the sole test for identifying female patients with chlamydial or gonococcal infections may miss some infected individuals.

Performance estimates for urine specimens are based on evaluation of urine obtained from the first part of the urine stream; performance on midstream collections has not been determined.
This assay **does** detect plasmid-free variants of *Chlamydia trachomatis*.

This assay **does not** detect *C pneumoniae*.

This assay has not been shown to cross-react with commensal (nonpathogenic) *Neisseria* species in the oropharynx.

**Clinical Reference**


**Performance**

**Method Description**

The HOLOGIC APTIMA Combo 2 Assay combines the technologies of target capture, transcription-mediated amplification, and dual kinetic assay. The detection of the rRNA amplification product sequences (amplicon) is achieved using nucleic acid hybridization. Single-stranded chemiluminescent DNA probes are labeled and combine with amplicon to form stable RNA:DNA hybrids. Light emitted from the labeled RNA:DNA hybrids is measured as photon signals in a luminometer. (Package insert: HOLOGIC APTIMA Combo 2 Assay 501798 002 2017-03)

**PDF Report**

No

**Day(s) and Time(s) Test Performed**

Monday through Saturday; Varies

**Analytic Time**

1 day

**Maximum Laboratory Time**

4 days

**Specimen Retention Time**

7 days

**Performing Laboratory Location**

Rochester
Fees and Codes

Fees
- Authorized users can sign in to Test Prices for detailed fee information.
- Clients without access to Test Prices can contact Customer Service 24 hours a day, seven days a week.
- Prospective clients should contact their Regional Manager. For assistance, contact Customer Service.

Test Classification
This test has been cleared, approved or is exempt by the U.S. Food and Drug Administration and is used per manufacturer's instructions. Performance characteristics were verified by Mayo Clinic in a manner consistent with CLIA requirements.

CPT Code Information
87491-Chlamydia trachomatis
87591-Neisseria gonorrhoeae

LOINC® Information

<table>
<thead>
<tr>
<th>Test ID</th>
<th>Test Order Name</th>
<th>Order LOINC Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>CGRNA</td>
<td>Chlamydia/Gonorrhoeae Amplified RNA</td>
<td>64017-7</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Result ID</th>
<th>Test Result Name</th>
<th>Result LOINC Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>SRC14</td>
<td>SOURCE:</td>
<td>31208-2</td>
</tr>
<tr>
<td>SRC17</td>
<td>SOURCE:</td>
<td>31208-2</td>
</tr>
<tr>
<td>34875</td>
<td>Neisseria gonorrhoeae amplified RNA</td>
<td>43305-2</td>
</tr>
<tr>
<td>34874</td>
<td>Chlamydia trachomatis amplified RNA</td>
<td>43304-5</td>
</tr>
</tbody>
</table>