Overview

Useful For
Detection of *Chlamydia trachomatis*

Method Name
TranscriptionMediatedAmplification

NY State Available
Yes

Specimen

Specimen Type
Varies

Necessary Information
Specimen source is required.

Specimen Required
Submit only 1 of the following specimens:

**Swab specimen must be collected** using an Aptima Collection Unisex Swab (T583), or Aptima Collection Multitest Swab (T584, formerly called Aptima Vaginal Swab Specimen Collection Kit). These swabs are contained in the Aptima Collection Kit.

**Supplies:** Swab, Aptima Male/Female Collection (T583)

**Specimen Type:** Endocervix

**Container/Tube:** Aptima Collection Unisex Swab (T583)

**Specimen Volume:** Swab

Collection Instructions:

1. Use cleaning swab (white shaft) to remove excess mucus from endocervix and discard.

2. Insert second swab (blue shaft) 1 to 1.5 cm into endocervical canal, and rotate swab gently for 30 seconds. Avoid touching vaginal wall when removing swab.

3. Place second swab into transport tube provided in collection kit. Snap off swab at score line so swab fits into closed tube.

4. Cap tube securely, and label tube with patient's entire name, and date and time of collection.

5. Transport and store swab container at 2 to 30 degrees C (refrigerate is preferred temperature) within 60 days of collection. If longer storage is needed, freeze at -20 to -70 degrees C for 12 months.
Test Definition: CTRNA
Chlamydia trachomatis Amplified RNA

Supplies: Swab, Aptima Multitest Swab Specimen Collection Kit (T584)

Specimen Type: Vaginal

Container/Tube: Aptima Collection Multitest Swab (T584)

Specimen Volume: Swab

Collection Instructions:

1. Insert swab (pink shaft) about 5 cm past introitus and rotate gently for 30 seconds.
2. Place swab into transport tube provided in collection kit. Snap off swab at score line so swab fits into closed tube.
3. Cap tube securely, and label tube with patient's entire name, and date and time of collection.
4. Transport and store swab container at 2 to 30 degrees C (refrigerate is preferred temperature) within 60 days of collection. If longer storage is needed, freeze at -20 to -70 degrees C for 12 months.

Supplies: Swab, Aptima Male/Female Collection (T583)

Specimen Type: Urethra (Males Only)

Container/Tube: Aptima Collection Unisex Swab (T583)

Specimen Volume: Swab

Collection Instructions:

1. Patient should not have urinated for at least 1 hour prior to collection.
2. With a rotating movement, insert swab (blue shaft) 2 to 4 cm into urethra.
3. Once inserted, rotate swab gently at least 1 full rotation using sufficient pressure to ensure swab comes into contact with all urethral surfaces. Allow swab to remain inserted for 2 to 3 seconds.
4. Place swab in transport tube provided in collection kit. Snap off swab at score line so swab fits into closed tube.
5. Cap tube securely, and label tube with patient's entire name, and date and time of collection.
6. Transport and store swab container at 2 to 30 degrees C (refrigerate is preferred temperature) within 60 days of collection. If longer storage is needed, freeze at -20 to -70 degrees C for 12 months.

Supplies: Aptima Urine Transport Tube (T582)

Specimen Type: Urine (Males and Females)

Container/Tube: Aptima Urine Specimen Transport Tube (T582)

Specimen Volume: 15-20 mL
**Test Definition: CTRNA**  
Chlamydia trachomatis Amplified RNA

**Collection Instructions:**

1. Patient should not have urinated for at least 1 hour prior to specimen collection.

2. Patient should collect first portion of random voided urine (first part of stream) into a sterile, plastic, preservative-free container.

3. Transfer 2 mL of urine into the urine specimen transport tube using the disposable pipette provided within 24 hours of collection. The correct volume of urine has been added when the fluid level is between the black fill lines on the urine transport tube. Place the labels on the transport tube so the black fill lines are still visible for volume confirmation at Mayo Clinic Laboratories.

4. Transport and store urine specimen transport container at 2 to 30 degrees C (refrigerate is preferred temperature) within 30 days of collection. If longer storage is needed, freeze at -20 to -70 degrees C for 12 months.

**Supplies:** Aptima Thin Prep Transport Tube (T652)

**Specimen Type:** ThinPrep Specimen (Endocervix)

**Container/Tube:** ThinPrep (also called PreservCyt) Collection Kit

**Specimen Volume:** 1 mL

**Collection Instructions:**

1. Aliquot ThinPrep specimen for *Chlamydia* and/or *Neisseria* testing before processing for Pap smear. For each specimen, use a new pair of clean gloves.

2. Vortex ThinPrep/PreservCyt vial 3 to 10 seconds. Within 1 minute of vortexing:
   a. Transfer 1 mL of specimen into the Aptima Specimen Transfer Tube (T652) using a disposable transfer pipette or a pipette tip containing a filter (aerosol barrier or hydrophobic plug).
   b. Process only 1 ThinPrep and transfer tube set at a time.
   c. Recap Aptima Specimen Transfer Tube tightly and gently invert 3 times to mix.

3. Label Aptima transfer tube with appropriate label.

4. Use remainder of ThinPrep specimen for Pap testing.

5. Transport and store specimen transfer container at 2 to 30 degrees C (refrigerate is preferred temperature) within 60 days of collection. If longer storage is needed, freeze at -20 to -70 degrees C for 12 months.

**Forms**

If not ordering electronically, complete, print, and send a [Microbiology Test Request](#) (T244) with the specimen.

**Specimen Minimum Volume**

Endocervix in PreservCyt: 1mL  
Urine: 2 mL  
Swabs (Endocervical, Urethral, Vaginal): Entire Collection
Reject Due To

<table>
<thead>
<tr>
<th>Other</th>
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<tbody>
<tr>
<td>Midstream urine specimen Overfilled or underfilled urine transport tubes Specimen collected into a SurePath device Transport tubes containing a cleaning swab or more than 1 swab</td>
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Specimen Stability Information

<table>
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<tr>
<th>Specimen Type</th>
<th>Temperature</th>
<th>Time</th>
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<tbody>
<tr>
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<td>APTIMA VIAL</td>
</tr>
<tr>
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<td>APTIMA VIAL</td>
</tr>
<tr>
<td></td>
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Clinical and Interpretive

Clinical Information

Chlamydia is caused by the obligate intracellular bacterium *Chlamydia trachomatis* and is the most prevalent sexually transmitted bacterial infection (STI) in the United States.\(^{(1,2)}\) In 2010, 1.3 million documented cases were reported to the Centers for Disease Control and Prevention (CDC).\(^{(2)}\) Given that 3 out of 4 infected women and 1 out of 2 infected men will be asymptomatic initially, the actual prevalence of disease is thought to be much greater than reported. The organism causes genitourinary infections in women and men and may be associated with dysuria and vaginal, urethral, or rectal discharge. In women, complications include pelvic inflammatory disease, salpingitis, and infertility. Approximately 25% to 30% of women who develop acute salpingitis become infertile.\(^{(2)}\) Complications among men are rare, but include epididymitis and sterility. Rarely, genital chlamydial infection can cause arthritis with associated skin lesions and ocular inflammation (Reiter syndrome). *C. trachomatis* can be transmitted from the mother during delivery and is associated with conjunctivitis and pneumonia. Finally, *C. trachomatis* may cause hepatitis and pharyngitis in adults.

Once detected, the infection is easily treated by a short course of antibiotic therapy. Annual chlamydia screening is now recommended for all sexually active women age 25 years and younger, and for older women with risk factors for infection, such as a new sex partner or multiple sex partners. The CDC also recommends that all pregnant women be given a screening test for *Chlamydia* infection.\(^{(2)}\) Repeat testing for test-of-cure is not recommended after treatment with a standard treatment regimen unless patient compliance is in question, reinfection is suspected, or the patient's symptoms persist. Repeat testing of pregnant women, 3 weeks after completion of therapy, is also recommended to ensure therapeutic cure.\(^{(2)}\)

Culture was previously considered to be the gold standard test for diagnosis of *C. trachomatis* infection.\(^{(2)}\) However, organisms are labile in vitro, and precise specimen collection, transportation, and processing conditions are required to maintain organism viability, which is necessary for successful culturing. In comparison, nucleic acid amplification testing (NAAT) provides superior sensitivity and specificity and is now the recommended method for diagnosis in most cases.\(^{(3-5)}\) Immunoassays and nonamplification DNA tests are also available for *C. trachomatis* detection, but these methods are significantly less sensitive and less specific than NAATs.\(^{(2)}\)

Improved screening rates and increased sensitivity of NAAT testing have resulted in an increased number of accurately diagnosed cases.\(^{(2)}\) Improved detection rates result from both the increased performance of the assay. Early identification of infection enables sexual partners to seek testing and/or treatment as soon as possible and
reduces the risk of disease spread. Prompt treatment reduces the risk of infertility in women.

**Reference Values**

**Negative**

**Interpretation**

A positive result indicates the presence of rRNA *Chlamydia trachomatis*.

A negative result indicates that rRNA for *C trachomatis* was not detected in the specimen.

The predictive value of an assay depends on the prevalence of the disease in any particular population. In settings with a high prevalence of sexually transmitted disease, positive assay results have a high likelihood of being true-positives. In settings with a low prevalence of sexually transmitted disease, or in any setting in which a patient's clinical signs and symptoms or risk factors are inconsistent with chlamydial or gonococcal urogenital infection, positive results should be carefully assessed and the patient retested by other methods, if appropriate.

**Cautions**

Care must be taken to avoid cross-contamination during handling of PreservCyt solution liquid Pap specimens. If testing PreservCyt specimens processed with the ThinPrep 2000 processor, it is important to follow procedures to reduce the risk for cross-contamination during Pap processing such as bleaching of the PreservCyt filter cap and changing gloves between each sample. Refer to the ThinPrep 2000 Processor Operator's Manual and the Aptima specimen for more guidance.

The performance of endocervical, vaginal, and male urethral swab specimens, male and female urine specimens, and PreservCyt solution liquid Pap specimens has not been evaluated in adolescents less than 16 years of age. The performance of vaginal swab specimens has not been evaluated in pregnant women.

This report is intended for use in clinical monitoring or management of patients; it is not intended for use in medico-legal applications.

Appropriate specimen collection and handling is necessary for optimal assay performance.

Results should be interpreted in conjunction with other laboratory and clinical information.

A negative test result does not exclude the possibility of infection. Improper specimen collection, concurrent antibiotic therapy, presence of inhibitors, or low numbers of organisms in the specimen (ie, below the sensitivity of the test) may cause false-negative test results.

In low prevalence populations, positive results must be interpreted carefully as false-positive results may occur more frequently than true-positive results in this setting.

In general, this assay should not be used to assess therapeutic success or failure, since nucleic acids from these organisms may persist for 3 weeks or more following antimicrobial therapy.

The presence of mucous does not interfere with this assay. However, this test requires endocervical cells, and if excess mucous is not removed prior to collection, adequate numbers of these cells may not be obtained.

No interference is expected in swab specimens due to:

- Blood
- Lubricants and spermicides
Test Definition: CTRNA
Chlamydia trachomatis Amplified RNA

The effects of use of tampons, douching, specimen types other than those listed in Specimen Required, and specimen collection variables have not been determined.

Testing of urine specimens with this method is not intended to replace cervical exam and endocervical sampling for diagnosis of urogenital infection; infections may result from other causes or concurrent infections may occur.

Testing urine specimens as the sole test for identifying female patients with chlamydial infections may miss some infected individuals.

Performance estimates for urine specimens are based on evaluation of urine obtained from the first part of the urine stream; performance on midstream collections has not been determined.

This assay does detect plasmid-free variants of Chlamydia trachomatis.

This assay does not detect C. pneumoniae.

Clinical Reference


Performance

Method Description
The HOLOGIC APTIMA Combo 2 Assay combines the technologies of target capture, transcription-mediated amplification, and dual kinetic assay. The detection of the rRNA amplification product sequences (amplicon) is achieved using nucleic acid hybridization. Single-stranded chemiluminescent DNA probes are labeled and combine with amplicon to form stable RNA:DNA hybrids. Light emitted from the labeled RNA:DNA hybrids is measured as photon signals in a luminometer. (Package insert: HOLOGIC APTIMA Combo 2 Assay 501798 Rev 002 2017-03)

PDF Report
No

Day(s) and Time(s) Test Performed
Monday through Saturday; Varies

Analytic Time
Test Definition: CTRNA
Chlamydia trachomatis Amplified RNA

1 day

Maximum Laboratory Time
3 days

Specimen Retention Time
7 days

Performing Laboratory Location
Rochester

Fees and Codes

Fees
- Authorized users can sign in to Test Prices for detailed fee information.
- Clients without access to Test Prices can contact Customer Service 24 hours a day, seven days a week.
- Prospective clients should contact their Regional Manager. For assistance, contact Customer Service.

Test Classification
This test has been cleared or approved by the U.S. Food and Drug Administration and is used per manufacturer’s instructions. Performance characteristics were verified by Mayo Clinic in a manner consistent with CLIA requirements.

CPT Code Information
87491

LOINC® Information

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<td>34874</td>
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