

## Overview

### Useful For

Aiding in the diagnosis of tularemia caused by *Francisella tularensis*

These assays should **not be used** as a test of cure as they are not quantitative and patients may remain seropositive for months to years following resolution of disease.

### Highlights

This test detects IgM and IgG class antibodies to *Francisella tularensis* in serum and may be used as an aid for the diagnosis of tularemia.

Serologic testing should be performed alongside other diagnostic methods, including culture of appropriate specimens. (**Note:** please notify microbiology laboratory in cases of suspected *F tularensis* to minimize exposure risk to bench technologists)

### Profile Information

Test ID	Reporting Name	Available Separately	Always Performed
TULM	F. tularensis Ab, IgM ELISA, S	No	Yes
TULG	F. tularensis Ab, IgG ELISA, S	No	Yes
TULI	F. tularensis Interpretation	No	Yes

### Method Name

Enzyme-Linked Immunosorbent Assay (ELISA)

### NY State Available

Yes

## Specimen

### Specimen Type

Serum

### Specimen Required

**Supplies:** Aliquot Tube, 5 mL (T465)

### Collection Container/Tube:

**Preferred:** Serum gel

**Acceptable:** Red top

**Submission Container/Tube:** Plastic vial

**Specimen Volume:** 0.6 mL

**Collection Instructions:** Centrifuge and aliquot serum.

**Specimen Minimum Volume**

0.5 mL

**Reject Due To**

Gross hemolysis	Reject
Gross lipemia	Reject
Gross icterus	Reject
Other	Heat inactivated specimen

**Specimen Stability Information**

Specimen Type	Temperature	Time	Special Container
Serum	Refrigerated (preferred)	7 days	
	Frozen	30 days	

**Clinical and Interpretive**

**Clinical Information**

*Francisella tularensis* is a small, intracellular, coccobacillary Gram negative bacterium and is an obligate pathogen in animals and humans, primarily maintained in rabbits, hares, cats, ticks and deerflies. *F. tularensis* is found throughout North America and parts of Asia, and similar to *Brucella* species is considered a potential agent of bioterrorism. Human infection with *F. tularensis* usually occurs through inhalation of infected aerosols, ingestion of contaminated meat or water, handling of diseased or sick animals, or through the bite of an infected arthropod (eg, tick, deerflies).

Following a 3 to 5 day incubation period, the clinical manifestations of infection with *F. tularensis* differ primarily depending on the site and route of infection. The most common form of disease is ulceroglandular (45%-80% of cases), which is associated with an arthropod (or animal) bite or another cause of skin barrier compromise. This leads to development of a painful papule which ultimately ulcerates following which the bacterium enters the lymphatic system. Glandular tularemia is similar in presentation to ulceroglandular disease, however it lacks the ulceration and more frequently causes septicemia. Other, less frequent clinical manifestations include oculoglandular (Parinaud syndrome), oropharyngeal and gastrointestinal disease, pneumonic or typhoidal tularemia.

Diagnostic testing options for *F. tularensis* primarily include culture and serology. Physicians suspecting tularemia should collect appropriate specimens (eg, skin lesion biopsy, lymph node aspirates, etc.) promptly and send for culture. The microbiology laboratory should be alerted to the possibility of *F. tularensis* to ensure that appropriate safety measures are taken to protect the laboratory technologists. Growth on culture is a definitive means of making a diagnosis of tularemia. Serologic testing may be used to support a diagnosis of current or recent tularemia in patients who are IgM positive, or seroconvert to IgM, or IgG positive in paired sera collected 2 to 3 weeks apart.

**Reference Values**

Negative

Reference values apply to all ages.

### Interpretation

IgM Result	IgG Result	Interpretation
Negative	Negative	No antibodies to <i>Francisella tularensis</i> detected. Antibody response may be negative in samples collected too soon following infection/exposure. Repeat testing on a new sample if clinically indicated.
Positive	Negative	IgM class antibodies to <i>F tularensis</i> detected, suggesting current or recent infection. Repeat testing in 2 to 3 weeks to detect seroconversion of IgG may be considered to confirm the diagnosis.
Positive	Borderline	
Borderline	Negative	Questionable presence of IgM antibodies to <i>F tularensis</i> . Consider repeat testing in 1 to 2 weeks.
Borderline	Positive	IgG class antibodies to <i>F tularensis</i> detected suggesting recent or past infection. Clinical correlation alongside presentation, exposure history and other laboratory findings required.
Borderline	Borderline	Questionable presence of IgM and IgG class antibodies to <i>F tularensis</i> . Consider repeat testing in 1 to 2 weeks.
Positive	Positive	IgM and IgG class antibodies to <i>F tularensis</i> detected suggesting current, recent or past infection. Clinical correlation alongside presentation, exposure history and other laboratory findings required.
Negative	Positive	IgG class antibodies to <i>F tularensis</i> detected suggesting recent or past infection. Clinical correlation alongside presentation, exposure history and other laboratory findings required.
Negative	Borderline	Questionable presence of IgG antibodies to <i>F tularensis</i> . Consider repeat testing in 1 to 2 weeks.

### Cautions

False negative results may occur in samples collected too soon following symptom onset, prior to the development of a detectable immune response. Repeat testing on new samples collected 2 to 4 weeks later may be helpful.

False positive results may occur in patients previously or currently infected with *Brucella* species. Other less frequent causes of cross-reactivity that have been reported include prior infection with *Yersinia*, *Salmonella*, or *Legionella* species.

IgM-class antibodies may be detectable as soon as 1 week after symptom onset and may remain detectable for multiple years following resolution of disease in some individuals. Therefore, an IgM positive result may not indicate current or recent infection in some cases.

There are multiple subspecies of *Francisella tularensis*, including *F tularensis* subspecies *tularensis*, *F tularensis* subspecies *holarctica* and *F tularensis* subspecies *novicida*, that are found throughout the northern hemisphere, including in the United States. The IgM and IgG anti-*F tularensis* ELISA tests used at Mayo Clinic Laboratories are based on the lipopolysaccharide (LPS) antigen of *F tularensis*. Although not directly tested, previous studies indicate that there are no antigenic differences between the LPS of *F tularensis* subspecies *tularensis* and the other subspecies. Therefore, these assays should not be used to differentiate between infection with the various *F tularensis* subspecies.

### Clinical Reference

1. Petersen JM, Schriefer ME, Araj GE: *Francisella* and *Brucella*. In Manual of Clinical Microbiology. 12th Edition. 2019
2. Nigrovic LE, Wingerter SL: [Tularemia](#). Infect Dis Clin North Am. 2008; 22(3): 489-504. doi: 10.1016/j.idc.2008.03.004

### Performance

#### Method Description

The ELISA is an immunoassay, which is particularly suited to the determination of antibodies in various kinds of samples. The reaction is based on the specific interaction of antibodies with their corresponding antigen. The test strips of the microtiter plate are coated with specific antigens of the pathogen of interest. If antibodies in the sample are present, they bind to the fixed antigen. A secondary antibody, which has been conjugated with the enzyme alkaline phosphatase, detects and binds to the immune complex. The colorless substrate p-nitrophenylphosphate is then converted into the colored product p-nitrophenol. The signal intensity of this reaction product is proportional to the concentration of the analyte in the sample and is measured photometrically. (Package insert: *Francisella tularensis* IgG/IgM ELISA, Immuno-Biological Laboratories Inc., Minneapolis, MN. Version No. 142.4)

#### PDF Report

No

#### Day(s) and Time(s) Test Performed

Tuesday and Thursday; 2 p.m.

#### Analytic Time

Same day/1 day

#### Maximum Laboratory Time

7 days

#### Specimen Retention Time

14 days

#### Performing Laboratory Location

Rochester

**Fees and Codes****Fees**

- Authorized users can sign in to [Test Prices](#) for detailed fee information.
- Clients without access to Test Prices can contact [Customer Service](#) 24 hours a day, seven days a week.
- Prospective clients should contact their Regional Manager. For assistance, contact [Customer Service](#).

**Test Classification**

This test has been cleared or approved by the U.S. Food and Drug Administration and is used per manufacturer's instructions. Performance characteristics were verified by Mayo Clinic in a manner consistent with CLIA requirements.

**CPT Code Information**

86668 x 2

**LOINC® Information**

Test ID	Test Order Name	Order LOINC Value
TULAB	F. tularensis Ab, IgM/IgG ELISA, S	93715-1

Result ID	Test Result Name	Result LOINC Value
TULM	F. tularensis Ab, IgM ELISA, S	93716-9
TULG	F. tularensis Ab, IgG ELISA, S	93717-7
TULI	F. tularensis Interpretation	93718-5