

## Overview

### Useful For

Detecting bacteria responsible for infections of sterile body fluids, tissues, or wounds

Determining the in vitro antimicrobial susceptibility of potentially pathogenic aerobic bacteria, if appropriate

This test is **not intended for** medicolegal use.

### Reflex Tests

Test ID	Reporting Name	Available Separately	Always Performed
COMM	Identification Commercial Kit	No, (Bill Only)	No
RMALD	Ident by MALDI-TOF mass spec	No, (Bill Only)	No
GID	Bacteria Identification	No, (Bill Only)	No
ISAE	Aerobe Ident by Sequencing	No, (Bill Only)	No
REFID	Additional Identification Procedure	No, (Bill Only)	No
SALS	Serologic Agglut Method 1 Ident	No, (Bill Only)	No
EC	Serologic Agglut Method 2 Ident	No, (Bill Only)	No
SHIG	Serologic Agglut Method 3 Ident	No, (Bill Only)	No
STAP	Identification Staphylococcus	No, (Bill Only)	No
STRP	Identification Streptococcus	No, (Bill Only)	No
BLA	Beta Lactamase	No, (Bill Only)	No
MIC	Sensitivity, MIC	No, (Bill Only)	No
SUS	Susceptibility	No, (Bill Only)	No
SIDC	Ident Serologic Agglut Method 4	No, (Bill Only)	No
PCRID	Identification by PCR	No, (Bill Only)	No

### Testing Algorithm

When this test is ordered, the reflex tests may be performed and charged. Antimicrobial agent appropriate to the organism and specimen source will be tested according to Mayo's practice and the laboratory's standard operating procedures.

See Special Instructions to review tables that provide a listing of the antimicrobials routinely tested in our laboratory

as well as antimicrobials that may be tested upon request. These tables are organized by isolate groups and are not all inclusive. Call 800-533-1710 and ask to speak to the Bacteriology Antimicrobial Susceptibility Testing Laboratory if the organism or antimicrobial of interest are not listed in these tables.

### Special Instructions

- [Aerobic Gram-Negative Bacilli Antimicrobials](#)
- [Additional Gram-Negative Bacteria Antimicrobials](#)
- [Staphylococcus, Enterococcus, Bacillus, and Related Genera Antimicrobials](#)
- [Additional Gram-Positive Bacteria Antimicrobials](#)

### Method Name

Conventional Culture Technique with Minimum Inhibitory Concentration (MIC) by Agar Dilution (if appropriate)

### NY State Available

Yes

### Specimen

#### Specimen Type

Varies

#### Shipping Instructions

**Specimen must arrive within 24 hours of collection.**

#### Necessary Information

**Specimen source is required;** include the specific anatomic source. Indicate whether it is a "surface" or "deep/surgical" specimen. **Do not** label only as "wound."

#### Specimen Required

**Preferred:**

**Specimen Type:** Closed abscess

**Container/Tube:** Sterile container

**Specimen Volume:** Entire collection

**Collection Instructions:** Aspirate the abscess contents with a syringe.

**Specimen Stability Information:** Ambient 24 hours (preferred)/Refrigerated 24 hours

**Acceptable:**

**Specimen Type:** Open abscess, swab, tissue, or fluid

**Supplies:** Culturette (BBL Culture Swab) (T092)

**Sources:** Abscess, aspirate, lesion, or wound

**Container/Tube:** Sterile container or culture transport swab (Dacron or rayon swab with aluminum or plastic)

shaft with either Stuart or Amies liquid medium)

**Collection Instructions:** For most open lesions and abscesses, remove superficial flora by decontaminating skin before collecting a specimen from advancing margin or base.

**Additional Information:**

1. If submitting a specimen from a source contaminated with usual flora, send at refrigerated temperature.
2. Refrigerated specimens are not suitable for isolation of *Neisseria* species.

**Specimen Stability Information:** Ambient 24 hours (preferred)/Refrigerated 24 hours

**Forms**

If not ordering electronically, complete, print, and send a [Microbiology Test Request](#) (T244) with the specimen.

**Specimen Minimum Volume**

0.5 mL

**Reject Due To**

Other	Specimen >24 hours Dry swab Frozen
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**Specimen Stability Information**

Specimen Type	Temperature	Time	Special Container
Varies	Ambient (preferred)	24 hours	
	Refrigerated	24 hours	

**Clinical and Interpretive**

**Clinical Information**

Sterile Body Fluids and Normally Sterile Tissues:

In response to infection, fluid may accumulate in any body cavity.

Wound, Abscess, Exudates:

Skin and soft tissue infections can occur as a result of a break in the skin surface, or they can occur as complications of surgery, trauma, human, animal, or insect bites or diseases that interrupt a mucosal or skin surface. Specimen collection is of utmost importance for these specimen types. For most open lesions and abscesses, remove the superficial flora by decontaminating the skin before collecting a specimen from the advancing margin or base. A closed abscess is the specimen site of choice. Aspirate the abscess contents with a syringe.

The specific anatomic site is required to establish possible contaminating flora in the area of specimen collection for appropriate reporting of culture results. For this reason, specimens should be labeled as to the specific anatomic source and to distinguish between "surface" and "deep/surgical" specimens. Do not label only as "wound."

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Antimicrobial susceptibility testing should be performed on pure culture isolates of pathogenic (or potentially pathogenic in special situations) bacteria grown from specimens that have been appropriately collected so as not to confuse clinically significant isolates with normal flora.

Antimicrobial susceptibility testing determines the minimum inhibitory concentration (MIC) value of selected antimicrobial agents against isolated potentially pathogenic bacteria. The MIC is the lowest antimicrobial concentration (of a series of increasing concentrations) that inhibits growth of the bacterium. Agar dilution MIC testing is performed by testing for growth of bacteria on agar plates containing varying concentrations of antimicrobial agents.

For each organism-antimicrobial agent combination, the Clinical and Laboratory Standards Institute provides interpretive criteria for determining whether the MIC should be interpreted as susceptible, susceptible dose dependent, intermediate, nonsusceptible, resistant, or Epidemiological Cutoff Value (ECV).

### Reference Values

No growth or usual flora

Identification of probable pathogens

Results are reported as minimal inhibitory concentration (MIC) in mcg/mL. Breakpoints (also known as "clinical breakpoints") are used to categorize an organism as susceptible, susceptible-dose dependent, intermediate, resistant, or nonsusceptible according to the Clinical and Laboratory Standards Institute (CLSI) guidelines.

In some instances an interpretive category cannot be provided based on available data and the following comment will be included: "There are no established interpretive guidelines for agents reported without interpretations."

Susceptible (S):

A category defined by a breakpoint that implies that isolates with an MIC at or below the susceptible breakpoint are inhibited by the usually achievable concentrations of antimicrobial agent when the dosage recommended to treat the site of infection is used, resulting in likely clinical efficacy.

Susceptible-Dose Dependent (SDD):

A category defined by a breakpoint that implies that susceptibility of an isolate depends on the dosing regimen that is used in the patient. In order to achieve levels that are likely to be clinically effective against isolates for which the susceptibility testing results are in the SDD category, it is necessary to use a dosing regimen (ie, higher doses, more frequent doses, or both) that results in higher drug exposure than that achieved with the dose that was used to establish the susceptible breakpoint. Consideration should be given to the maximum approved literature-supported dosage regimens, because higher exposure gives the highest probability of adequate coverage of a SDD isolate. The drug label should be consulted for recommended doses and adjustment for organ function.

Intermediate (I):

A category defined by a breakpoint that includes isolates with MICs within the intermediate range that approach usually attainable blood and tissue levels and for which response rates may be lower than for susceptible isolates.

**Note:** The intermediate category implies clinical efficacy in body sites where the drugs are physiologically concentrated or when a higher than normal dosage of a drug can be used. This category also includes a buffer zone, which should prevent small, uncontrolled, technical factors from causing major discrepancies in interpretations, especially for drugs with narrow pharmacotoxicity margins.

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**Resistant (R):**

A category defined by a breakpoint that implies that isolates with an MIC at or above the resistant breakpoint are not inhibited by the usually achievable concentrations of the agent with normal dosage schedules and/or that demonstrate MICs that fall in the range in which specific microbial resistance mechanisms are likely, and clinical efficacy of the agent against the isolate has not been reliably shown in treatment studies.

**Nonsusceptible (NS):**

A category used for isolates for which only a susceptible breakpoint is designated because of the absence or rare occurrence of resistant strains. Isolates for which the antimicrobial agent MICs are above the value indicated for the susceptible breakpoint should be reported as nonsusceptible.

**Note:** An isolate that is interpreted as nonsusceptible does not necessarily mean that the isolate has a resistance mechanism. It is possible that isolates with MICs above the susceptible breakpoint that lack resistance mechanisms may be encountered within the wild-type distribution subsequent to the time the susceptible-only breakpoint was set.

**Epidemiological Cutoff Value (ECV):**

The MIC that separates microbial populations into those with and without acquired resistance (non-wild-type or wild-type, respectively). The ECV defines the highest MIC for the wild type population of isolates. ECVs are based on in vitro data only, using MIC distributions. ECVs are **not** clinical breakpoints, and the clinical relevance of ECVs for a particular patient has not yet been identified or approved by CLSI or any regulatory agency.

When an ECV is reported, the following comment will be included: "This MIC is consistent with the Epidemiological Cutoff Value (ECV) observed in isolates [WITH / WITHOUT] acquired resistance; however, correlation with treatment outcome is unknown."

(CLSI. Performance Standards for Antimicrobial Susceptibility Testing. 29th edition. CLSI supplement M100. Wayne, PA: Clinical and Laboratory Standards Institute; 2019)

**Interpretation**

Any microorganism is considered significant and is reported when the anatomical source is considered sterile and no resident flora is expected. For specimens contaminated with the usual bacterial flora, bacteria that are potentially pathogenic are identified.

A "susceptible" category result and a low minimum inhibitory concentration value indicate in vitro susceptibility of the organism to the antimicrobial tested.

Refer to the Reference Values section for interpretation of various categories.

**Cautions**

When antimicrobial susceptibilities are performed, in vitro susceptibility does not guarantee clinical response. Therefore, the decision to treat with a particular agent should not be based solely on the antimicrobial susceptibility testing result.

**Clinical Reference**

1. Forbes BA, Sahm DF, Weissfeld AS: Infections of the urinary tract. In Bailey and Scott's Diagnostic Microbiology. 12th edition. St. Louis, MO, Mosby, 2007, pp 842-855

2. Cockerill FR: Conventional and genetic laboratory tests used to guide antimicrobial therapy. Mayo Clin Proc 1998;73:1007-1021

3. Procop GW, Church DL, Hall GS, et al: Chapter 2, Introduction to Microbiology Part II: Guidelines for the Collection, Transport, Processing, Analysis, and Reporting of Cultures From Specific Specimen Sources. In Koneman's Color Atlas and Textbook of Diagnostic Microbiology. 7th Edition. Wolters Kluwer Health, . 2017. pp 66-110

4. CLSI: Performance Standards for Antimicrobial Susceptibility Testing. 29th edition. CLSI supplement M100. Wayne, PA: Clinical and Laboratory Standards Institute; 2019, pp 3-5, 246

## Performance

### Method Description

Specimens are cultured to enriched and/or selective media appropriate to the anatomic location and the scope of microorganisms expected. Cultures are incubated for 3 to 14 days depending on the specimen source. Pathogens or possible pathogens are identified using one or a combination of the following techniques: commercial identification strips or panels, matrix-assisted laser desorption/ionization time-of-flight (MALDI-TOF) mass spectrometry, conventional biochemical tests, carbon source utilization, real-time polymerase chain reaction (PCR), and nucleic acid sequencing of the 16S ribosomal RNA (rRNA) gene. "Usual flora" is reported as such (as appropriate to the specimen). (Clinical Microbiology Procedures Handbook. Edited by AL Leber. Vol 1. Fourth edition. Washington DC, ASM Press, 2016. Sections 3.5, 3.7, 3.9, 3.10, 3.13)

When antimicrobial susceptibility testing is performed, an agar dilution method is used for routine testing. The antimicrobial is added to agar in various concentrations depending upon levels attainable in serum, urine, or both. A standardized suspension of the organism is applied to the agar plates, which are incubated for 16 to 18 hours at 35 degrees C. Complete inhibition of all but one colony or a very fine residual haze represents the end-point. (Clinical and Laboratory Standards Institute: Methods for Dilution Antimicrobial Susceptibility Tests for Bacteria That Grow Aerobically. 11th edition. CLSI standard M07. Wayne, PA, 2018)

### PDF Report

No

### Day(s) and Time(s) Test Performed

[Monday through Sunday](#)

### Analytic Time

5 days

### Maximum Laboratory Time

14 days

### Specimen Retention Time

2 days

### Performing Laboratory Location

Rochester

## Fees and Codes

### Fees

- Authorized users can sign in to [Test Prices](#) for detailed fee information.
- Clients without access to Test Prices can contact [Customer Service](#) 24 hours a day, seven days a week.

- Prospective clients should contact their Regional Manager. For assistance, contact [Customer Service](#).

**Test Classification**

This test has been cleared, approved or is exempt by the U.S. Food and Drug Administration and is used per manufacturer's instructions. Performance characteristics were verified by Mayo Clinic in a manner consistent with CLIA requirements.

**CPT Code Information**

- 87070-Bacterial Culture, Aerobic
- 87186-Antimicrobial Susceptibility, Aerobic Bacteria, MIC-per organism for routine battery (if appropriate)
- 87187-Susceptibility per drug and per organism for drugs not in routine battery (if appropriate)
- 87077-Identification commercial kit (if appropriate)
- 87077-Ident by MALDI-TOF mass spec (if appropriate)
- 87077-Bacteria Identification (if appropriate)
- 87077-Additional Identification procedure (if appropriate)
- 87077-Identification Staphylococcus (if appropriate)
- 87077-Identification Streptococcus (if appropriate)
- 87147 x 1-3-Serologic agglut method 1 ident (if appropriate)
- 87147-Serologic agglut method 2 ident (if appropriate)
- 87147 x 4-Serologic agglut method 3 ident (if appropriate)
- 87147 x 2-6-Serologic Agglut Method 4 Ident (if appropriate)
- 87153-Aerobe ident by sequencing (if appropriate)
- 87185-Beta lactamase (if appropriate)
- 87798-Identification by PCR (if appropriate)

**LOINC® Information**

Test ID	Test Order Name	Order LOINC Value
GENS	Bacterial Culture, Aerobic + Susc	634-6

Result ID	Test Result Name	Result LOINC Value
GENS	Bacterial Culture, Aerobic + Susc	634-6