Overview

Useful For
Detecting the deletion or addition of the SRY gene in conjunction with conventional chromosome studies (CHRCB / Chromosome Analysis, Congenital Disorders, Blood)

Genetics Test Information
This test is appropriate to aid in detecting the presence or absence of the SRY gene in XX males and XY females. Testing must be ordered in conjunction with conventional chromosome studies (CHRCB / Chromosome Analysis, Congenital Disorders, Blood).

Reflex Tests

<table>
<thead>
<tr>
<th>Test ID</th>
<th>Reporting Name</th>
<th>Available Separately</th>
<th>Always Performed</th>
</tr>
</thead>
<tbody>
<tr>
<td>_I099</td>
<td>Interphases, 25-99</td>
<td>No, (Bill Only)</td>
<td>No</td>
</tr>
<tr>
<td>_I300</td>
<td>Interphases, &gt;=100</td>
<td>No, (Bill Only)</td>
<td>No</td>
</tr>
<tr>
<td>_IL25</td>
<td>Interphases</td>
<td>No, (Bill Only)</td>
<td>No</td>
</tr>
<tr>
<td>_ML10</td>
<td>Metaphases, &gt;=10</td>
<td>No, (Bill Only)</td>
<td>No</td>
</tr>
<tr>
<td>_PADD</td>
<td>Probe, +1</td>
<td>No, (Bill Only)</td>
<td>No</td>
</tr>
<tr>
<td>_PB02</td>
<td>Probe, +2</td>
<td>No, (Bill Only)</td>
<td>No</td>
</tr>
<tr>
<td>_PB03</td>
<td>Probe, +3</td>
<td>No, (Bill Only)</td>
<td>No</td>
</tr>
<tr>
<td>_PB1</td>
<td>Probe Set, 1st</td>
<td>No, (Bill Only)</td>
<td>No</td>
</tr>
</tbody>
</table>

Testing Algorithm
This test includes a charge for application of the first probe set (2 FISH probes) and professional interpretation of results. Additional charges will be incurred for application of all reflex probes performed. Analysis charges will be incurred based on the number of cells analyzed per probe set. If no cells are available for analysis, no analysis charges will be incurred.

Special Instructions
- Final Disposition of Fetal/Stillborn Remains
- Informed Consent for Genetic Testing
- Informed Consent for Genetic Testing (Spanish)

Method Name
Fluorescence In Situ Hybridization (FISH)

NY State Available
Yes

Specimen

Specimen Type
Test Definition: SRYF
SRY, Yp11.3, FISH

Varies

**Additional Testing Requirements**
This test must be ordered in conjunction with conventional chromosome studies (CHRCB / Chromosome Analysis, Congenital Disorders, Blood).

**Shipping Instructions**
Advise Express Mail or equivalent if not on courier service.

**Necessary Information**
Provide a reason for referral with each specimen. The laboratory will not reject testing if this information is not provided, but appropriate testing and interpretation may be compromised or delayed.

**Specimen Required**
Submit only 1 of the following specimens:

**Supplies:** Refrigerate/Ambient Shipping Box, 5 lb (T329)

**Specimen Type:** Amniotic fluid

**Container/Tube:** Amniotic fluid container

**Specimen Volume:** 20-25 mL

**Collection Instructions:**
1. Optimal timing for specimen collection is during 14 to 18 weeks of gestation, but specimens collected at other weeks of gestation are also accepted. Provide gestational age at the time of amniocentesis.

2. Discard the first 2 mL of amniotic fluid.

3. Place the tubes in a Styrofoam container (T329).

4. Fill remaining space with packing material.

**Additional Information:**
1. Unavoidably, about 1% to 2% of mailed-in specimens are not viable.

2. Bloody specimens are undesirable.

3. If the specimen does not grow in culture, you will be notified within 7 days of receipt.

4. Results will be reported and also telephoned or faxed, if requested.

**Supplies:** Hank's Solution (T132)

**Specimen Type:** Autopsy

**Container/Tube:** Sterile container with sterile Hank's balanced salt solution (T132), Ringer's solution, or normal saline
**Specimen Volume:** 4 mm diameter

**Collection Instructions:**

1. Wash biopsy site with an antiseptic soap.
2. Thoroughly rinse area with sterile water.
3. Do not use alcohol or iodine preparations.
4. Biopsy specimens are best taken by punch biopsy to include full thickness of dermis.

**Specimen Type:** Blood

**Container/Tube:** Green top (sodium heparin)

**Specimen Volume:** 5 mL

**Collection Instructions:**

1. Invert several times to mix blood.
2. Other anticoagulants are not recommended and are harmful to the viability of the cells.

**Supplies:** CVS Media (RPMI) and Small Dish (T095)

**Specimen Type:** Chorionic villus

**Container/Tube:** 15-mL tube containing 15 mL of transport media

**Specimen Volume:** 20-25 mg

**Collection Instructions:**

1. Collect specimen by the transabdominal or transcervical method.
2. Transfer chorionic villi to a Petri dish containing transport medium (T095).
3. Using a stereomicroscope and sterile forceps, assess the quality and quantity of the villi and remove any blood clots and maternal decidua.

**Specimen Type:** Fixed cell pellet

**Container/Tube:** Sterile container with a 3:1 fixative (methanol:glacial acetic acid)

**Specimen Volume:** Entire specimen

**Supplies:** Hank’s Solution (T132)

**Specimen Type:** Products of conception or stillbirth

**Container/Tube:**
Sterile container with sterile Hank's balanced salt solution (T132), Ringer's solution, or normal saline

**Specimen Volume:** 1 cm\(^3\) of placenta (including 20-mg of chorionic villi) and a 1-cm\(^3\) biopsy specimen of muscle/fascia from the thigh

**Collection Instructions:** If a fetus cannot be specifically identified, collect villus material or tissue that appears to be of fetal origin.

**Additional Information:** Do not send entire fetus.

**Supplies:** Hank's Solution (T132)

**Specimen Type:** Skin biopsy

**Container/Tube:** Sterile container with sterile Hank's balanced salt solution (T132), Ringer's solution, or normal saline

**Specimen Volume:** 4 mm diameter

**Collection Instructions:**

1. Wash biopsy site with an antiseptic soap.
2. Thoroughly rinse area with sterile water.
3. Do not use alcohol or iodine preparations.
4. A local anesthetic may be used.
5. Biopsy specimens are best taken by punch biopsy to include full thickness of dermis.

**Forms**

1. **New York Clients-Informed consent is required.** Document on the request form or electronic order that a copy is on file. The following documents are available in Special Instructions:

   - [Informed Consent for Genetic Testing](#) (T576)
   - [Informed Consent for Genetic Testing-Spanish](#) (T826)

2. **Final Disposition of Fetal/Stillborn Remains** (if fetal specimen is sent) in Special Instructions.

**Specimen Minimum Volume**

- Amniotic Fluid: 5 mL
- Autopsy, Skin Biopsy: 4 mm
- Blood: 2 mL
- Chorionic Villi: 5 mg
- Fixed Cell Pellet: 1 pellet
- Products of Conception: 1 cm\(^3\)

**Reject Due To**

All specimens will be evaluated at Mayo Clinic Laboratories for test suitability.
Specimen Stability Information

<table>
<thead>
<tr>
<th>Specimen Type</th>
<th>Temperature</th>
<th>Time</th>
<th>Special Container</th>
</tr>
</thead>
<tbody>
<tr>
<td>Varies</td>
<td>Refrigerated (preferred)</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>Ambient</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Clinical and Interpretive

Clinical Information

This test is appropriate for individuals with a 46,XX karyotype and phenotypically normal male external genitalia, a 46,XY karyotype and phenotypically normal female external genitalia, clinical features suggestive of 46,XX testicular disorder of sex development with normal male external genitalia, and clinical features suggestive of 46,XY complete gonadal dysgenesis.

The SRY (sex-determining region on the Y chromosome) gene is required for normal embryonic wolffian (male) genital development, although numerous other genes are involved in completing the process of normal male development. Some gene mutations block the action of SRY in development. Thus, a 46,XY individual with an SRY deletion or mutation will develop as a female, and a 46,XX individual with translocation of SRY to 1 X chromosome will develop as a male. Structural abnormalities of the Y chromosome result in a spectrum of abnormalities from primary infertility (male or female) to various forms of ambiguous genitalia. SRY-negative 46,XX males often have ambiguous genitalia, whereas those who are positive for SRY usually have a normal male phenotype with azoospermia. SRY-negative 46,XY females may have another mutation, such as 1 involving the SOX9 gene.

We recommend conventional chromosome studies (CHRCB / Chromosome Analysis, Congenital Disorders, Blood) to detect Y chromosome abnormalities and to rule out other chromosome abnormalities or translocations, and FISH studies to detect cryptic translocations involving the SRY region that are not demonstrated by conventional chromosome studies.

Reference Values

An interpretive report will be provided.

Interpretation

Any male individual with an SRY signal on a structurally normal Y chromosome is considered negative for a deletion in the region tested by this probe. Any patient with a FISH signal pattern indicating loss of the critical region will be reported as having a deletion of the regions tested by this probe. Any patient with a FISH signal on an X chromosome will be reported as having a cryptic X;Y translocation involving the critical region.

Cautions

Because this FISH test is not approved by the U.S. Food and Drug Administration, it is important to confirm SRY deletions/duplications by other established methods, such as clinical history or physical evaluation.

Chromosomal microarray (CMAC / Chromosomal Microarray, Congenital, Blood or CMAP / Chromosomal Microarray, Prenatal, Amniotic Fluid/Chorionic Villus Sampling) may be the more appropriate test to detect unbalanced translocations, deletions or duplications. A

Interfering factors
-Cell lysis caused by forcing the blood quickly through the needle

-Use of an improper anticoagulant or improperly mixing the blood with the anticoagulant

-Excessive transport time

-Inadequate amount of specimen may not permit adequate analysis

-Improper packaging may result in broken, leaky, and contaminated specimen during transport

-Exposure of the specimen to temperature extremes (freezing or >30 degrees C) may kill cells and interfere with attempts to culture cells

-In prenatal specimens, a bloody specimen may interfere with attempts to culture cells and contamination by maternal cells may cause interpretive problems

Supportive Data

Using a probe for the SRY critical region, FISH analysis was performed on a series of 46 patient specimens, peripheral blood or amniotic fluid, and results were compared to cytogenetic analyses and the patient's phenotype.

Of 20 phenotypic females:
-12 with a 45,X karyotype or an X duplication exhibited no SRY signal
-8 with a 46,XY karyotype or an abnormal Y were SRY positive

Of 20 phenotypic males:
-7 of 8 with a 46,XX karyotype were SRY negative

-13 with a 46, XY with a normal or rearranged Y chromosome were SRY positive

Of 25 controls:
-13 males exhibited SRY on the Y chromosome
-12 females exhibited no SRY signal

Clinical Reference


Performance

Method Description
Test Definition: SRYF
SRY, Yp11.3, FISH

This test is performed using a commercially available enumeration strategy probe set including SRY (Yp11.3) and X chromosome control probe (DXZ1). Metaphase cells are examined for the presence of SRY.(Unpublished Mayo method)

PDF Report
No

Day(s) and Time(s) Test Performed
Specimens are processed Monday through Sunday.

Results reported Monday through Friday, 8 a.m. to 5 p.m.

Analytic Time
7 days

Maximum Laboratory Time
10 days

Specimen Retention Time
Amniotic Fl. (remaining supernatant/whole fluid aliquots): Discarded 14 days after report. Blood: 4 weeks. Products of Conception (identifiable fetal tissue): Cremated quarterly after results reported. All Other Specimens: Discarded when results reported.

Performing Laboratory Location
Rochester

Fees and Codes

Fees
- Authorized users can sign in to Test Prices for detailed fee information.
- Clients without access to Test Prices can contact Customer Service 24 hours a day, seven days a week.
- Prospective clients should contact their Regional Manager. For assistance, contact Customer Service.

Test Classification
This test was developed using an analyte specific reagent. Its performance characteristics were determined by Mayo Clinic in a manner consistent with CLIA requirements. This test has not been cleared or approved by the U.S. Food and Drug Administration.

CPT Code Information
88271x2, 88291-DNA probe, each (first probe set), Interpretation and report

88271x2-DNA probe, each; each additional probe set (if appropriate)

88271x1-DNA probe, each; coverage for sets containing 3 probes (if appropriate)

88271x2-DNA probe, each; coverage for sets containing 4 probes (if appropriate)

88271x3-DNA probe, each; coverage for sets containing 5 probes (if appropriate)

88273 w/modifier 52-Chromosomal in situ hybridization, less than 10 cells (if appropriate)
Test Definition: SRYF
SRY, Yp11.3, FISH

88273-Chromosomal in situ hybridization, 10-30 cells (if appropriate)

88274 w/modifier 52-Interphase in situ hybridization, <25 cells, each probe set (if appropriate)

88274-Interphase in situ hybridization, 25 to 99 cells, each probe set (if appropriate)

88275-Interphase in situ hybridization, 100 to 300 cells, each probe set (if appropriate)

LOINC® Information

<table>
<thead>
<tr>
<th>Test ID</th>
<th>Test Order Name</th>
<th>Order LOINC Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>SRYF</td>
<td>SRY, Yp11.3, FISH</td>
<td>81748-6</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Result ID</th>
<th>Test Result Name</th>
<th>Result LOINC Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>52003</td>
<td>Result Summary</td>
<td>50397-9</td>
</tr>
<tr>
<td>52005</td>
<td>Interpretation</td>
<td>69965-2</td>
</tr>
<tr>
<td>54565</td>
<td>Result</td>
<td>62356-1</td>
</tr>
<tr>
<td>CG717</td>
<td>Reason for Referral</td>
<td>42349-1</td>
</tr>
<tr>
<td>CG718</td>
<td>Specimen</td>
<td>31208-2</td>
</tr>
<tr>
<td>52006</td>
<td>Source</td>
<td>31208-2</td>
</tr>
<tr>
<td>52007</td>
<td>Method</td>
<td>49549-9</td>
</tr>
<tr>
<td>52004</td>
<td>Additional Information</td>
<td>48767-8</td>
</tr>
<tr>
<td>53850</td>
<td>Disclaimer</td>
<td>62364-5</td>
</tr>
<tr>
<td>52008</td>
<td>Released By</td>
<td>18771-6</td>
</tr>
</tbody>
</table>