

Overview

Useful For

Evaluation of pediatric blood specimens for chromosomal abnormalities associated with hematologic malignancies for diagnostic and prognostic purposes in patients being considered for enrollment in Children's Oncology Group (COG) clinical trials and research protocols

Highlights

Cytogenetics testing is important for the diagnostic and prognostic classification of pediatric neoplasia and it is a critical element for enrollment of children into clinical trials affiliated with the Children's Oncology Group (COG). For over 25 years the Mayo Clinic Genomics Laboratory has served as one of a select number of laboratories in the United States approved by the COG for the conventional chromosome analysis and FISH analysis of pediatric bone marrow, peripheral blood, and tissue specimens.

All enrollment-required elements of cytogenetics testing will be electronically submitted by the Mayo Clinic Genomics laboratory within the guidelines of COG.

Reflex Tests

Test ID	Reporting Name	Available Separately	Always Performed
ML20C	COG Metaphases, 1-19	No, (Bill Only)	No
M25C	COG Metaphases, 20-25	No, (Bill Only)	No
MG25C	COG Metaphases, >25	No, (Bill Only)	No
_STAC	Ag-Nor/CBL Stain	No, (Bill Only)	No

Testing Algorithm

This test is only performed on specimens from pediatric patients who are candidates for enrollment in Children's Oncology Group (COG) clinical trials and research protocols.

This test includes a charge for cell culture of fresh specimens and professional interpretation of results. Analysis charges will be incurred for total work performed, and generally include 2 banded karyograms and the analysis of 20 metaphase cells. If no metaphase cells are available for analysis, no analysis charges will be incurred. If additional analysis work is required, additional charges may be incurred.

-If this test is ordered and the laboratory is informed that the patient is not on a COG protocol, this test will be canceled and automatically reordered as CHRHB / Chromosome Analysis, Hematologic Disorders, Blood.

Method Name

Cell Culture without Mitogens followed by Chromosome Analysis

NY State Available

No

Specimen

Specimen Type

Whole blood

Advisory Information

This test is only performed on specimens from pediatric patients being considered for enrollment in a Children's Oncology Group (COG) protocol. For all other patients, order CHRHB / Chromosomes, Hematologic Disorders, Blood.

For children in whom disease relapse or a secondary myeloid neoplasm is a concern and enrollment in a new COG protocol is being considered; order COGBM / COG, Chromosome Analysis, Hematologic Disorders, Bone Marrow.

Consultation with personnel from the Genomics Laboratory is recommended when considering blood studies for hematologic disorders. Call 800-533-1710 and ask for the Cytogenetics Genetic Counselor on call.

Shipping Instructions

Advise Express Mail or equivalent if not on courier service.

Necessary Information

1. Provide a reason for referral with each specimen, as well as flow cytometry and/or a bone marrow pathology report and a Children's Oncology Group (COG) protocol number. The laboratory will not reject testing if this information is not provided, but appropriate testing and interpretation may be compromised or delayed.
2. If a child has received an opposite sex bone marrow transplant prior to specimen collection for this protocol, convey this information to the laboratory.

Specimen Required

Container/Tube: Green top (sodium heparin)

Specimen Volume: 6 mL

Collection Instructions:

Invert several times to mix blood.

Specimen Minimum Volume

3 mL

Reject Due To

All specimens will be evaluated at Mayo Clinic Laboratories for test suitability.

Specimen Stability Information

Specimen Type	Temperature	Time	Special Container
Whole blood	Ambient (preferred)		
	Refrigerated		

Clinical and Interpretive

Clinical Information

Clonal chromosomal abnormalities play a central role in the pathogenesis, diagnosis, and treatment monitoring of pediatric hematologic malignancies. Whenever possible, it is best to do chromosome studies for neoplastic hematologic disorders on bone marrow. Bone marrow studies are more sensitive and the chances of finding metaphases are about 95%, compared with only a 60% chance for blood studies. When it is not possible to collect bone marrow, chromosome studies on blood may be useful.

When blood cells are cultured in a medium without mitogens, the observation of any chromosomally abnormal clone may be consistent with a neoplastic process.

Characteristic chromosome rearrangements and copy number patterns may help classify a pediatric hematologic malignancy. For example, t(1;19)(q23;p13.3) is typically observed in B-lymphoblastic leukemia/lymphoma (B-ALL) and t(8;21)(q22;q22) defines a specific subset of patients with acute myeloid leukemia; while t(7;14)(q35;q11.2) is associated with T-lymphoblastic leukemia/lymphoma. Confirmation of classic gene fusions associated with the above translocations together with evaluation for other recurrent abnormalities are available within the appropriate COG FISH panels; COGBF / B-Lymphoblastic Leukemia/Lymphoma, Children's Oncology Group Enrollment Testing, FISH, Varies; COGTF / T-Cell Acute Lymphoblastic Leukemia (T-ALL), Children's Oncology Group Enrollment Testing, FISH, Varies; and COGMF / Acute Myeloid Leukemia (AML), Children's Oncology Group Enrollment Testing, FISH, Varies. Some rearrangements identified by chromosomal analysis may be extremely rare but are known, recurrent entities for which the Mayo Clinic Genomics Laboratory has the most extensive catalogue of FISH testing to confirm the corollary gene fusions.

Metaphase FISH confirmation of classic translocations which are cryptic and not visually detectable by chromosome analysis [ie, t(12;21)] associated with ETV6/RUNX1 fusion) is performed as required by COG and is included as part of the electronic case submission by the Mayo Clinic Genomics Laboratory to COG for central review.

Additional cytogenetic techniques such as chromosomal microarray (CMAH / Chromosomal Microarray, Hematologic Disorders, Varies) may be helpful to resolve questions related to ploidy (hyperdiploid clone vs doubled hypodiploid clone) or to resolve certain clonal structural rearrangements such as the presence or absence of intrachromosomal amplification of chromosome 21 (iAMP21). Occasionally, characterization of balanced or complex rearrangements presumed to involve critical genes (such as a kinase activating gene and a fusion partner) may be characterizable by Mate Pair sequencing (MTRBL / MatePair, Targeted Rearrangements, Congenital, Blood); a clinically validated next-generation sequencing technique developed at Mayo Clinic.

Reference Values

An interpretative report will be provided.

Interpretation

[The presence of an abnormal clone usually indicates a malignant neoplastic process.](#)

The absence of an apparent abnormal clone in blood may result from a lack of circulating abnormal cells and not from an absence of disease.

On rare occasions, the presence of an abnormality may be associated with a congenital abnormality and, thus, not related to a malignant process. When this situation is suspected, follow-up with a medical genetics consultation is recommended.

Cautions

Bone marrow specimens are preferred over peripheral blood specimens for the diagnosis of neoplastic hematologic disorders. When peripheral blood must be used, FISH studies may detect some disorders better than conventional chromosome studies.

-FISH studies also may be superior for other hematological disorders when compared to conventional chromosome studies utilizing blood specimens.

Interfering factors:

Technical:

- Cell lysis caused by forcing blood quickly through the needle at collection
- Use of an improper anticoagulant (sodium heparin is best) or improperly mixing the blood with the anticoagulant
- Excessive transport time
- Exposure of the specimen to temperature

Biological:

- Abnormalities missed due to sampling error
- Subtle structural chromosome abnormalities may not be detected by conventional chromosome analysis
- Neoplastic cells not dividing or not circulating in the bloodstream

Clinical Reference

1. Dewald GW, Ketterling RP, Wyatt WA, Stupca PJ: Cytogenetic studies in neoplastic hematologic disorders. In Clinical Laboratory Medicine, Second edition. Edited by KD McClatchey. Baltimore, Williams and Wilkins, 2002, pp 658-685
2. Rigolin GM, Cibien F, Martinelli S, et al: Chromosome aberrations detected by conventional karyotyping using novel mitogens in chronic lymphocytic leukemia with "normal" FISH: correlations with clinicobiological parameters. Blood 2012 Mar 8;119(10):2310-2313
3. Swerdlow SH, Campo E, Harris NL, et al.: International Agency for Research on Cancer (IARC): World Health Organization (WHO) classification of tumours of haematopoietic and lymphoid tissues. IARC Press, Oxford University Press, 2017
4. Arber DA, Borowitz MJ, Cessna M, et al: Initial Diagnostic Workup of Acute Leukemia: Guideline From the College of American Pathologists and the American Society of Hematology. Arch Pathol Lab Med 2017 Oct;141(10):1342-1393

Performance

Method Description

[A cell count is performed to establish a plating volume. Based on the cell count, a corresponding volume of blood is added to 2 culture flasks containing culture medium and incubated for 24 to 48 hours at 37 degrees C. In the harvest process, the cells are exposed to colcemid and hypotonic solution, and are fixed with glacial acetic acid and methanol. Metaphase cells are dropped onto microscope slides and are stained by G-banding. Other staining methods are employed as needed. Twenty metaphases are usually examined. If a clone is suspected, but not confirmed within 20 metaphases, 30 metaphases will be analyzed. Minimal evidence for the presence of an abnormal clone is defined as 2 or more metaphases with the same structural abnormality or chromosome gain \(trisomy\), or 3 or more metaphases lacking the same chromosome. All cells analyzed are captured using a](#)

[computerized imaging system, and 1 or more karyograms from each clone are prepared to document the abnormality and to permit systematic interpretation of the anomalies.\(Unpublished Mayo method\)](#)

PDF Report

No

Day(s) and Time(s) Test Performed

Specimens are processed Monday through Sunday.

Results reported Monday through Friday, 8 a.m.-5 p.m.

Analytic Time

9 days

Maximum Laboratory Time

11 days

Specimen Retention Time

4 weeks

Performing Laboratory Location

Rochester

Fees and Codes**Fees**

- Authorized users can sign in to [Test Prices](#) for detailed fee information.
- Clients without access to Test Prices can contact [Customer Service](#) 24 hours a day, seven days a week.
- Prospective clients should contact their Regional Manager. For assistance, contact [Customer Service](#).

Test Classification

This test was developed and its performance characteristics determined by Mayo Clinic in a manner consistent with CLIA requirements. This test has not been cleared or approved by the U.S. Food and Drug Administration.

CPT Code Information

88237, 88291-Tissue culture for neoplastic disorders; bone marrow, blood, Interpretation and report

88264 w/ modifier 52-Chromosome analysis with less than 20 cells (if appropriate)

88264-Chromosome analysis with 20 to 25 cells (if appropriate)

88264, 88285-Chromosome analysis with greater than 25 cells (if appropriate)

88283-Additional specialized banding technique (if appropriate)

LOINC® Information

Test ID	Test Order Name	Order LOINC Value
COGBL	COG-Chromosomes, Hematologic, Blood	In Process



Test ID	Test Order Name	Order LOINC Value
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Result ID	Test Result Name	Result LOINC Value
602315	Result Summary	50397-9
602316	Interpretation	69965-2
602317	Result	62356-1
GC024	Reason for Referral	42349-1
602318	Specimen	31208-2
602319	Source	31208-2
602320	Method	85069-3
602321	Banding Method	62359-5
602322	Additional Information	48767-8
602323	Released By	18771-6