

Hemophilia A F8 Gene, Intron 1 Inversion Known Mutation Analysis, Prenatal, Varies

### Overview

#### **Useful For**

Prenatal testing for hemophilia A when a F8 intron 1 inversion has been identified in a family member.

### **Reflex Tests**

Test Id	Reporting Name	Available Separately	Always Performed
CULAF	Amniotic Fluid	Yes	No
	Culture/Genetic Test		
CULFB	Fibroblast Culture for	Yes	No
	Genetic Test		
MATCC	Maternal Cell	Yes	No
	Contamination, B		
_STR1	Comp Analysis using STR	No, (Bill only)	No
	(Bill only)		
_STR2	Add'l comp analysis w/STR	No, (Bill only)	No
	(Bill Only)		

### **Genetics Test Information**

Detects the intron 1 inversion in the F8 gene. The intron 1 inversion mutation accounts for approximately 5% of mutations associated with severe hemophilia A.

Intron 1 inversion known mutation analysis on a prenatal specimen can only be performed when there is a known intron 1 inversion in the family.

# **Testing Algorithm**

If amniotic fluid is received, amniotic fluid culture/genetic test will be added and charged separately. If chorionic villus specimen is received, fibroblast culture for genetic test will be added and charged separately. For any prenatal specimen that is received, maternal cell contamination studies will be added. A maternal whole blood sample is required to perform this test.

The following algorithms are available in Special Instructions:

- -Hemophilia Carrier Testing Algorithm
- -Hemophilia Testing Algorithm

# **Special Instructions**

- Informed Consent for Genetic Testing
- Hemophilia Carrier Testing Algorithm
- Hemophilia Testing Algorithm
- Hemophilia A Patient Information
- Informed Consent for Genetic Testing (Spanish)



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### **Method Name**

Polymerase Chain Reaction (PCR)

#### NY State Available

Yes

# Specimen

# **Specimen Type**

Varies

# **Necessary Information**

<u>Hemophilia A Patient Information</u> is required, see Special Instructions. Testing may proceed without the patient information, however, the information aids in providing a more thorough interpretation. Ordering providers are strongly encouraged to fill out the form and send with the specimen.

### Specimen Required

Advise Express Mail or equivalent if not on courier service

Results will be reported and also telephoned or faxed, if requested.

Due to the complexity of prenatal testing, consultation with the laboratory is required for all prenatal testing. Prenatal specimens can be sent Monday through Thursday and must be received by 5 p.m. CST on Friday in order to be processed appropriately. All prenatal specimens must be accompanied by a maternal blood specimen. Order MATCC / Maternal Cell Contamination, Molecular Analysis on the maternal specimen.

# Submit only 1 of the following specimens:

Specimen Type: Amniotic fluid

Container/Tube: Amniotic fluid container

Specimen Volume: 5-10 mL Collection Instructions:

- 1. Optimal timing for specimen collection is during 14 to 18 weeks of gestation, but specimens collected at other weeks of gestation are also accepted.
- 2. Discard the first 2 mL of amniotic fluid. If the culture will be performed in conjunction with chromosome analysis and alpha-fetoprotein, a total of approximately 25 to 30 mL will be needed for the combined studies.

Specimen Stability Information: Ambient (preferred) <24 hours/Refrigerated

# **Additional Information:**

- 1. Place the tubes in a Styrofoam container (T329).
- 2. Fill remaining space with packing material.
- 3. Unavoidably, about 1% to 2% of mailed-in specimens are not viable.
- 4. Bloody specimens are undesirable.
- 5. If the specimen does not grow in culture, you will be notified within 7 days of receipt.



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Specimen Type: Chorionic villi

Container/Tube: 15-mL tube containing 15 mL of transport media

**Specimen Volume:** 20-30 mg **Collection Instructions:** 

1. Collect specimen by the transabdominal or transcervical method.

2. Transfer the chorionic villi specimen to a Petri dish containing transport medium (T095).

3. Using a stereomicroscope and sterile forceps, assess the quality and quantity of the villi and remove any blood clots

and maternal decidua.

Specimen Stability Information: Refrigerated (preferred) <24 hours/Ambient

Specimen Type: Confluent cultured cells

Container/Tube: T-25 flask

Specimen Volume: 2 Flasks approximately 90% confluent

**Collection Instructions:** Submit confluent cultured cells from another laboratory. **Specimen Stability Information:** Ambient (preferred) <24 hours/Refrigerated

**Additional Information:** There will be no culture charge.

#### **Forms**

1. New York Clients-Informed consent is required. Document on the request form or electronic order that a copy is on file. The following documents are available in Special Instructions:

-Informed Consent for Genetic Testing (T576)

-Informed Consent for Genetic Testing-Spanish (T826)

2. If not ordering electronically, complete, print, and send a Coagulation Test Request (T753) with the specimen.

# **Specimen Minimum Volume**

Amniotic fluid: 10 mL Chorionic villi: 5 mg

### **Reject Due To**

All specimens will be evaluated at Mayo Clinic Laboratories for test suitability.

### Specimen Stability Information

Specimen Type	Temperature	Time	Special Container
Varies	Varies		

# Clinical & Interpretive

# **Clinical Information**

Hemophilia A (HA) is caused by a deficiency of clotting factor VIII (FVIII). HA is an X-linked recessive bleeding disorder that affects approximately 1 in 5,000 males. Males are typically affected with bleeding symptoms, whereas carrier females generally do not have bleeding symptoms but are at risk of having affected sons. Rarely, approximately 10% of carrier females have FVIII activity levels below 35% and are at risk for bleeding.



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Bleeding is the most common clinical symptom in individuals with HA and correlates with FVIII activity levels. FVIII activity levels of <1% are associated with severe disease, 1% to 5% activity with moderate disease, and 5% to 40% with mild disease. In males with severe deficiency, spontaneous bleeding may occur. In individuals with mild HA, bleeding may occur only after surgery or trauma.

FVIII is encoded by the factor VIII (*F8*) gene. Approximately 98% of patients with a diagnosis of HA are found to have a mutation in *F8* (ie, intron 1 and 22 inversions, point mutations, insertions, and deletions). The intron 1 inversion mutation accounts for approximately 5% of mutations associated with severe HA. This inversion is typically not identified in patients with mild or moderate HA.

Intron 1 inversion known mutation analysis on a prenatal specimen can only be performed when there is a known intron 1 inversion in the family.

It is recommended that the F8 mutation be confirmed in the affected male or obligate carrier female prior to testing at-risk individuals. Affected males are identified by FVIII activity (F8A / Coagulation Factor VIII Activity Assay, Plasma) and clinical evaluation, while obligate carrier females are identified by family history assessment. Of note, not all females with an affected son are germline carriers of a F8 mutation, as de novo mutations in F8 do occur. Approximately 20% of mothers of isolated cases do not have an identifiable germline F8 mutation. Importantly, there is a small risk for recurrence even when the familial F8 mutation is not identified in the mother of the affected patient due to the possibility of germline mosaicism.

### **Reference Values**

Not applicable

# Interpretation

An interpretive report will be provided.

### **Cautions**

Obtaining a medical genetics or hematology (coagulation) consultation prior to ordering is advisable. Molecular genetic or hemophilia center consultation is available for all possible hemophilia A cases and is particularly indicated in complex cases or in situations in which the diagnosis is atypical or uncertain.

This assay detects only the F8 intron 1 inversion mutation. Thus, a negative result does not exclude the presence of other mutations in F8.

The intron 1 inversion mutation targeted by this assay is found in approximately 5% of individuals with severe hemophilia A; if an intron 1 inversion has not been already identified in the family, the assay may be uninformative.

Test results should be interpreted in the context of clinical findings, family history, and other laboratory data. Errors in our interpretation of results may occur if information given is inaccurate or incomplete.

### Clinical Reference

1. Antonarakis SE, Rossiter JP, Young M, et al: Factor VIII gene inversions in severe hemophilia A: results of an international consortium study. Blood 1995;86(6):2206-2212



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- 2. Rossiter JP, Young M, Kimberland ML, et al: Factor VIII gene inversions causing severe hemophilia A originate almost exclusively in male germ cells. Hum Mol Genet 1994;3(7):1035-1039
- 3. Castaldo G, D'Argenio V, Nardiello P, et al: Haemophilia A: molecular insights. Clin Chem Lab Med 2007;45(4):450-461
- 4. Pruthi RK: Hemophilia: A Practical Approach to Genetic Testing. Mayo Clin Proc 2005;80:1485-1499

### **Performance**

### **Method Description**

Genomic DNA from amniotic fluid or chorionic villi is amplified by PCR with primers specific for the F8 intron 1 inversion mutation. (Bagnall RD, Waseem N, Green PM, Giannelli F: Recurrent inversion breaking intron 1 of the factor VIII gene is a frequent cause of severe hemophilia A. Blood 2002;99[1]:168-174)

### **PDF Report**

No

# Day(s) Performed

Monday through Friday

### Report Available

28 to 35 days

# **Specimen Retention Time**

Extracted DNA indefinitely, patient must opt-out.

### **Performing Laboratory Location**

Rochester

# **Fees & Codes**

### **Fees**

- Authorized users can sign in to <u>Test Prices</u> for detailed fee information.
- Clients without access to Test Prices can contact <u>Customer Service</u> 24 hours a day, seven days a week.
- Prospective clients should contact their Regional Manager. For assistance, contact <u>Customer Service</u>.

# **Test Classification**

This test was developed, and its performance characteristics determined by Mayo Clinic in a manner consistent with CLIA requirements. This test has not been cleared or approved by the US Food and Drug Administration.

### **CPT Code Information**

81403



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# **LOINC®** Information

Test ID	Test Order Name	Order LOINC® Value
F81P	HA F8 Intron 1 Inversion, AF or CVS	82342-7

Result ID	Test Result Name	Result LOINC® Value
35138	HA F8 Int1 KM Reason for Referral	42349-1
35004	HA F8 Intron 1 Inversion, AF or CVS	82342-7
35005	F81P Interpretation	69047-9
35006	HA F8 Int1 KM Reviewed By	18771-6