Test Definition: NAIFA

Antinuclear Antibodies, HEp-2 Substrate, IgG, Serum

Overview

Useful For

Evaluation of patients suspected of having systemic autoimmune rheumatic disease (ANA-associated rheumatic diseases or connective tissue disease) or organ-specific autoimmune diseases such as autoimmune liver diseases

Testing Algorithm

For more information see:
-Antinuclear Antibody Interpretive Algorithm
-Connective Tissue Disease Cascade

Special Instructions

- Connective Tissue Disease Cascade

Method Name

Indirect Immunofluorescence

NY State Available

Yes

Specimen

Specimen Type

Serum

Specimen Required

Supplies: Sarstedt Aliquot Tube 5 mL (T914)
Collection Container/Tube:
Preferred: Serum gel
Acceptable: Red top
Submission Container/Tube: Plastic vial
Specimen Volume: 0.5 mL serum
Collection Instructions: Centrifuge and aliquot serum into a plastic vial.

Forms

If not ordering electronically, complete, print, and send General Test Request (T239) with the specimen.

Specimen Minimum Volume

0.3 mL

Reject Due To

Gross hemolysis	OK
Gross lipemia	OK
Gross icterus	OK

Specimen Stability Information

Specimen Type	Temperature	Time	Special Container
Serum	Refrigerated (preferred)	21 days	
	Frozen	28 days	

Clinical & Interpretive

Clinical Information

Autoantibodies targeting antigens in the nuclear region in the human epithelial type 2 (HEp-2) cell line substrate using the indirect immunofluorescence assay (IFA) have traditionally been called antinuclear antibody (ANA). ANA is the commonly performed antibody test in the initial evaluation of patients with systemic autoimmune rheumatic diseases (also referred to as connective tissue disease). Classic ANA-associated rheumatic diseases include systemic lupus erythematosus (SLE), mixed connective tissue disease (MCTD), Sjogren syndrome (Sjs), and systemic sclerosis (SSc) including CREST (calcinosis, Raynaud phenomenon, esophageal dysmotility, sclerodactyly, telangiectasia) and inflammatory myopathies (IM) such as dermatomyositis (DM).(1-4) Testing for ANA may also be of diagnostic relevance in the differential evaluation of autoimmune liver diseases (ALD).(5-6)

The classical ANA patterns (antibodies targeting the nuclear region) include homogeneous, speckled, centromere, nuclear dots, and nucleolar. These patterns are routinely reported by most clinical laboratories. SLE patients and those with SSc, Sjs, IM (such as anti-synthetase syndrome and necrotizing autoimmune myopathy) or ALD have also been shown to have clinically significant antibodies that react with antigens in other cellular compartments such as the cytoplasm and structures associated mitosis or mitotic patterns with HEp-2 substrate.(reviewed in 1-3) Based on the increasing recognition of these non-nuclear antigenic targets and their documented clinical relevance, the first International Consensus on ANA Patterns established a classification tree for ANA with alpha-numeric anti-cell code for each pattern with a recommendation for a change in terminology from antinuclear antibody to anticellular antibody.(2) These changes are relevant as in addition the nuclear patterns, the classification includes cytoplasmic and mitotic patterns with descriptions for their interpretation, associated antibody targets and clinical associations when available.(4)

The diagnosis of ANA-associated rheumatic diseases is usually based on a set of criteria of which the presence on anticellular antibody or specific associated antibodies may be components. Of all ANA-associated rheumatic diseases, the presence of anticellular antibody is considered mandatory entry criterion by the 2019 European League Against Rheumatism and the American College of Rheumatology classification criteria for SLE.(7) Since cytoplasmic staining patterns may be reported as "ANA negative" or as a comment with no quantitative or titer result, some patients with

clinicopathological symptoms consistent with neuropsychiatric SLE would not qualify for entry based on where testing is performed.(8-10) This limitation may therefore exclude patients who may meet the clinical and other laboratory criteria for disease but are not reported as "ANA positive" due to the use of the current terminology. In an international inception cohort of newly diagnosed SLE patients, 6.2% were anticellular antibody-negative with 1.5% testing positive for isolated cytoplasmic or mitotic pattern.(11) In addition, a recent investigation of various HEp-2 IFA kits showed variabilities in the expression of specific patterns with high reproducibilities between tests for centromere, multiple nuclear dots, nuclear coarse speckled, nuclear homogeneous, and cytoplasmic reticular antimitochondrial antibody patterns.(12)

Overall, the anticellular antibody is a screening test for ANA-associated rheumatic diseases with variable sensitivities in the different clinical subsets but lacks diagnostic specificity.(1-4) Therefore, positive results require confirmation with the use of specific ANA-associated antibody tests except for the centromere pattern which is very characteristic for patients with limited diffuse SSc. Confirmation of a positive anticellular antibody test result may be guided by HEp-2 IFA patterns and/or titer, patient's clinical presentation or in some cases the patient's demographic.(13)

Reference Values

<1:80 (Negative)

Interpretation

Presence of anticellular antibody (formerly antinuclear antibody) is a feature of systemic autoimmune rheumatic diseases such as systemic lupus erythematosus, mixed connective tissue disease, Sjogren syndrome and systemic sclerosis and some inflammatory myopathies (dermatomyositis, anti-synthetase syndrome and necrotizing autoimmune myopathy). It may also be of diagnostic relevance in patients with autoimmune liver diseases.

Patients’ sera are screened at 1:80. The following nuclear patterns and their titers are reported: centromere, homogeneous, nuclear dots, nucleolar, speckled, fine dense speckled (also referred to as DFS70), and proliferating cell nuclear antigen (PCNA). If observed, the following cytoplasmic patterns are reported: reticular/AMA (antimitochondrial antibody), cytoplasmic speckled, fibrillar, polar/Golgi-like, or rods and rings. The spindle fiber and centrosome mitotic patterns are also reported if observed. Reported patterns may help guide differential diagnosis, although they may not be specific for individual antibodies or diseases. Negative results do not necessarily rule out systemic autoimmune rheumatic disease.

Anticellular antibody test lacks diagnostic specificity and is associated with some cancers, infectious, and inflammatory conditions, with variable prevalence in healthy individuals. The lack of diagnostic specificity requires confirmation of positive results using associated antibody tests such as those targeting extractable nuclear antigens.

Cautions

Some patients without clinical evidence of systemic autoimmune rheumatic disease (SARD) may be positive for anticellular antibody. This occurs at variable prevalence depending on the patient demographics. A positive result may also precede clinical manifestation of SARD or be associated with some viral or chronic infections, cancers, or use of certain medications. All results must be reported in the appropriate clinical context as the performance of the test can be variable.

Clinical Reference

1. Agmon-Levin N, Damoiseaux J, Kallenberg C, et al. International recommendations for the assessment of

autoantibodies to cellular antigens referred to as anti-nuclear antibodies. Ann Rheum Dis. 2014;73(1):17-23
2. Chan EK, Damoiseaux J, Gabriel Carballo O, et al. Report of the First International Consensus on Standardized Nomenclature of Antinuclear Antibody HEp-2 Cell Patterns 2014-2015. Front Immunol. 2015;6:412
3. Bossuyt X, De Langhe E, Borghi MO, Meroni PL. Understanding and interpreting antinuclear antibody tests in systemic rheumatic diseases. Nat Rev Rheumatol. 2020;16(12):715-726
4. International Consensus on ANA Patterns. Nomenclature and Classification Tree. ICAP; 2021. Updated February 2025. Accessed October 31, 2025. Available at www.anapatterns.org/trees.php
5. European Association for the Study of the Liver. EASL Clinical Practice Guidelines: The diagnosis and management of patients with primary biliary cholangitis. J Hepatol. 2017;67(1):145-172
6. Younossi ZM, Bernstein D, Shiffman ML, et al. Diagnosis and management of primary biliary cholangitis. Am J Gastroenterol. 2019;114(1):48-63
7. Aringer M, Costenbader K, Daikh D, et al. 2019 European League Against Rheumatism/American College of Rheumatology classification criteria for systemic lupus erythematosus. Arthritis Rheumatol. 2019;71(9):1400-1412
8. Naides SJ, Genzen JR, Abel G, Bashleben C, Ansari MQ. Antinuclear antibodies testing method variability: A survey of participants in the College of American Pathologists' Proficiency Testing Program. J Rheumatol. 2020;47(12):1768-1773
9. Van Hoovels L, Broeders S, Chan EKL, et al. Current laboratory and clinical practices in reporting and interpreting anti-nuclear antibody indirect immunofluorescence (ANA IIF) patterns: results of an international survey. Auto Immun Highlights. 2020;11(1):17
10. Tebo AE, Schmidt RL, Kadkhoda K, et al. The antinuclear antibody HEp-2 indirect immunofluorescence assay: a survey of laboratory performance, pattern recognition and interpretation. Auto Immun Highlights. 2021;12(1):4
11. Choi MY, Clarke AE, ST Pierre Y, et al. Antinuclear antibody-negative systemic lupus erythematosus in an international inception cohort. Arthritis Care Res. 2019;71(1):893-902
12. Silva MJ, Dellavance A, Baldo DC, et al. Interkit reproducibility of the indirect immunofluorescence assay on HEp-2 cells depends on the immunofluorescence reactivity intensity and pattern. Front Immunol. 2022;12:798322
13. Nandjwada SL, Peterson LK, Mayes MD, et al. Ethnic differences in autoantibody diversity and hierarchy: More clues from a US cohort of patients with systemic sclerosis. J Rheumatol. 2016;43(10):1816-1824

Performance

Method Description

Antibodies to nuclear antigens in a human epithelial type 2 (HEp-2) cell line by an indirect immunofluorescent technique. Commercial slides prepared from HEp-2 cells are used as a substrate. IgG antibodies in serum specimens are detected after incubation of serum with the commercial slides by the addition of a fluorescein isothiocyante (FITC)-labeled antihuman-IgG reagent. All patient specimens are initially screened at 1:80.(Package insert: NOVA Lite DAPI ANA. Inova Diagnostics; 06/2018)

PDF Report

No

Day(s) Performed

Monday through Saturday

Report Available

2 to 3 days

Specimen Retention Time

14 days

Performing Laboratory Location

Mayo Clinic Laboratories - Rochester Superior Drive

Fees & Codes

Fees

- Authorized users can sign in to Test Prices for detailed fee information.
- Clients without access to Test Prices can contact Customer Service 24 hours a day, seven days a week.
- Prospective clients should contact their account representative. For assistance, contact Customer Service.

Test Classification

This test has been cleared, approved, or is exempt by the US Food and Drug Administration and is used per manufacturer's instructions. Performance characteristics were verified by Mayo Clinic in a manner consistent with CLIA requirements.

CPT Code Information

86039

LOINC® Information

Test ID	Test Order Name	Order LOINC® Value
NAIFA	Antinuclear Ab, HEp-2 Substrate, S	59069-5

Result ID	Test Result Name	Result LOINC® Value
ANAH	Antinuclear Ab, HEp-2 Substrate, S	59069-5
1TANA	ANA Titer:	33253-6
1PANA	ANA Pattern:	49311-4
2TANA	ANA Titer 2:	33253-6
2PANA	ANA Pattern 2:	49311-4
CYTQL	Cytoplasmic Pattern:	55171-3
LCOM	Lab Comment:	77202-0