

Angiosarcoma, MYC (8q24) Amplification, FISH, Tissue

#### Overview

#### **Useful For**

Identifying *MYC* amplification to aid in the differentiation of cutaneous angiosarcomas from atypical vascular lesions after radiotherapy

#### **Reflex Tests**

| Test Id | Reporting Name     | Available Separately | Always Performed |
|---------|--------------------|----------------------|------------------|
| _PBCT   | Probe, +2          | No, (Bill Only)      | No               |
| _PADD   | Probe, +1          | No, (Bill Only)      | No               |
| _PB02   | Probe, +2          | No, (Bill Only)      | No               |
| _PB03   | Probe, +3          | No, (Bill Only)      | No               |
| _IL25   | Interphases, <25   | No, (Bill Only)      | No               |
| _1099   | Interphases, 25-99 | No, (Bill Only)      | No               |
| _1300   | Interphases, >=100 | No, (Bill Only)      | No               |

## **Testing Algorithm**

This test does not include a pathology consultation. If a pathology consult is requested, PATHC / Pathology Consultation should be ordered, and the appropriate fluorescence in situ hybridization (FISH) test will be performed at an additional charge.

This test includes a charge for application of the first probe set (2 FISH probes) and professional interpretation of results.

Additional charges will be incurred for all reflex probes performed. Analysis charges will be incurred based on the number of cells analyzed per probe set. If no cells are available for analysis, no analysis charges will be incurred.

Appropriate ancillary probes may be performed at consultant discretion to render comprehensive assessment. Any additional probes will have the results included within the final report and will be performed at an additional charge.

#### **Method Name**

Fluorescence In Situ Hybridization (FISH)

## **NY State Available**

Yes

### Specimen

## **Specimen Type**

Tissue



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## **Ordering Guidance**

This test does not include a pathology consultation. If a pathology consultation is requested, order PATHC / Pathology Consultation, and appropriate testing will be added at the discretion of the pathologist and performed at an additional charge.

Multiple oncology (cancer) gene panels are also available. For more information see <u>Hematology, Oncology, and Hereditary Test Selection Guide</u>.

## **Additional Testing Requirements**

To resolve atypical fluorescence in situ hybridization results, confirmation testing by microarray testing is available; order CMAPT / Chromosomal Microarray, Tumor, Formalin-Fixed Paraffin-Embedded.

## **Shipping Instructions**

Advise Express Mail or equivalent if not on courier service.

## **Necessary Information**

- **1.** A pathology report is required for testing to be performed. If not provided, appropriate testing and interpretation may be compromised or delayed. Acceptable pathology reports include working drafts, preliminary pathology, or surgical pathology reports.
- 2. The following information must be included in the report provided:
- -Patient name
- -Block number must be on all blocks, slides, and paperwork
- -Date of collection
- -Tissue source
- **3.** A reason for testing must be provided. If this information is not provided, an appropriate indication for testing may be entered by Mayo Clinic Laboratories.

#### Specimen Required

Submit only 1 of the following specimens:

#### **Preferred:**

Specimen Type: Tissue block

**Collection Instructions:** Submit a formalin-fixed, paraffin-embedded tumor tissue block. Blocks prepared with alternative fixation methods will be attempted but are less favorable for successful results by fluorescence in situ hybridization testing; provide fixation method used.

#### **Additional Information:**

- 1. Paraffin-embedded specimens can be from any anatomic location (skin, soft tissue, lymph node, etc).
- 2. Bone specimens that have been decalcified will be attempted for testing, but the success rate is approximately 50%.

## Acceptable:

Specimen Type: Tissue slides

Slides: 1 Hematoxylin and eosin stained and 4 unstained

Collection Instructions: Submit 1 slide stained with hematoxylin and eosin and 4 consecutive unstained, positively



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charged, unbaked slides with 5 micron-thick sections of the tumor tissue.

#### **Forms**

If not ordering electronically, complete, print, and send an Oncology Test Request (T729) with the specimen.

#### **Specimen Minimum Volume**

Slides: 1 Hematoxylin and eosin stained and 2 unstained

#### **Reject Due To**

All specimens will be evaluated at Mayo Clinic Laboratories for test suitability.

#### **Specimen Stability Information**

| Specimen Type | Temperature         | Time | Special Container |
|---------------|---------------------|------|-------------------|
| Tissue        | Ambient (preferred) |      |                   |
|               | Refrigerated        |      |                   |

## **Clinical & Interpretive**

#### **Clinical Information**

Postradiation cutaneous angiosarcoma is a malignancy associated with very poor outcome and is consequently treated aggressively. Atypical vascular lesions are also associated with radiation therapy but are considered to be benign and do not require aggressive management. Therefore, the differentiation of these neoplasms is of considerable clinical importance. Postradiation cutaneous angiosarcomas often demonstrate high-level amplification of *MYC*, whereas reactive and benign vascular lesions do not show amplification of *MYC*.

#### **Reference Values**

An interpretive report will be provided.

## Interpretation

MYC will be clinically interpreted as positive, negative, or equivocal.

The MYC locus is reported as amplified when the MYC:D8Z2 ratio is 2.0 or greater and demonstrates 6 or more copies of MYC.

A MYC:D8Z2 ratio less than 2.0 or showing a ratio of 2.0 or greater with less than 6 copies of *MYC* is considered to lack amplification of *MYC*.

#### **Cautions**

This test is not approved by the US Food and Drug Administration, and it is best used as an adjunct to existing clinical and pathologic information.

This fluorescence in situ hybridization (FISH) assay does not rule out other chromosome abnormalities.



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Fixatives other than formalin (eg, Prefer, Bouin's) may not be successful for FISH assays. Non-formalin fixed specimens will not be rejected.

Paraffin-embedded tissues that have been decalcified may not be successful for FISH analysis. The success rate of FISH studies on decalcified tissue is approximately 50%, but FISH will be attempted if sufficient tumor is present for analysis.

FISH studies will be attempted if sufficient tumor is present for analysis. The pathologist reviewing the hematoxylin and eosin-stained slide may find it necessary to cancel testing if insufficient tissue/tumor is available for testing.

If no FISH signals are observed post hybridization, the case will be released indicating a lack of FISH results.

## **Supportive Data**

The probe set was independently validated in a blinded study on 23 paraffin-embedded primary and post radiation angiosarcoma tissue samples and 25 noncancerous control specimens. The normal controls were used to generate the normal cutoffs. *MYC* amplification was detected in 4 (17.4%) of the angiosarcomas and the incidence is consistent with published reports.

#### Clinical Reference

- 1. Mentzel T, Schildhaus H, Palmedo G, Buttner R, Kutzner H. Postradiation cutaneous angiosarcoma after treatment of breast carcinoma is characterized by *MYC* amplification in contrast to atypical vascular lesions after radiotherapy and control cases: clinicopathological, immunohistochemical and molecular analysis of 66 cases. Mod Pathol. 2012;25(1):75-85
- 2. Manner J, Radlwimmer B, Hohenberger P, et al. *MYC* high level gene amplification is a distinctive feature of angiosarcomas after irradiation or chronic lymphedema. Am J Pathol. 2010;176(1):34-39
- 3. WHO Classification of Tumours Editorial Board. Soft Tissue and Bone. 5th ed. IARC; 2020. World Health Organization Classification of Tumours. Vol 3

## **Performance**

## **Method Description**

This test is performed using a commercially available MYC probe set with a MYC probe and a chromosome 8 centromere probe (D8Z2). The selection of tissue and the identification of target areas on the hematoxylin and eosin (H and E)-stained slide are performed by a pathologist. Using the H and E-stained slide as a reference, target areas are etched with a diamond-tipped engraving tool on the back of the unstained slide to be assayed. The probe set is hybridized to the appropriate target areas, and 2 technologists each independently analyze 30 interphase nuclei (60 total) with the results expressed as a ratio of MYC:D8Z2 signals.(Unpublished Mayo method)

#### PDF Report

No

## Day(s) Performed

Monday through Friday



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## Report Available

7 to 10 days

#### **Specimen Retention Time**

Slides and H and E used for analysis are retained by the laboratory in accordance with regulatory requirements. Client provided paraffin blocks and extra unstained slides (if provided) will be returned after testing is complete.

### **Performing Laboratory Location**

Mayo Clinic Laboratories - Rochester Main Campus

#### **Fees & Codes**

### **Fees**

- Authorized users can sign in to <u>Test Prices</u> for detailed fee information.
- Clients without access to Test Prices can contact <u>Customer Service</u> 24 hours a day, seven days a week.
- Prospective clients should contact their account representative. For assistance, contact Customer Service.

#### **Test Classification**

This test was developed and its performance characteristics determined by Mayo Clinic in a manner consistent with CLIA requirements. It has not been cleared or approved by the US Food and Drug Administration.

#### **CPT Code Information**

88271x2, 88291-DNA probe, each (first probe set), Interpretation and report

88271x2-DNA probe, each; each additional probe set (if appropriate)

88271x1-DNA probe, each; coverage for sets containing 3 probes (if appropriate)

88271x2-DNA probe, each; coverage for sets containing 4 probes (if appropriate)

88271x3-DNA probe, each; coverage for sets containing 5 probes (if appropriate)

88274 w/modifier 52-Interphase in situ hybridization, <25 cells, each probe set (if appropriate)

88274-Interphase in situ hybridization, 25 to 99 cells, each probe set (if appropriate)

88275-Interphase in situ hybridization, 100 to 300 cells, each probe set (if appropriate)

## **LOINC®** Information

| Test ID | Test Order Name                    | Order LOINC® Value |
|---------|------------------------------------|--------------------|
| MASF    | MYC (8q24), Angiosarcoma, FISH, Ts | 101384-6           |

| Result ID | Test Result Name    | Result LOINC® Value |
|-----------|---------------------|---------------------|
| 54606     | Result Summary      | 50397-9             |
| 54609     | Interpretation      | 69965-2             |
| 54608     | Result              | 62356-1             |
| CG896     | Reason for Referral | 42349-1             |
| 54610     | Specimen            | 31208-2             |
| 54611     | Source              | 31208-2             |



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| 54612 | Tissue ID              | 80398-1 |
|-------|------------------------|---------|
| 54613 | Method                 | 85069-3 |
| 54614 | Released By            | 18771-6 |
| 55126 | Additional Information | 48767-8 |
| 53818 | Disclaimer             | 62364-5 |